
Anexos

Anexo I. Listado de publicaciones

Revistas

1. C.E. Rodríguez-Rodríguez, D. Lucas, E. Barón, P. Gago-Ferrero, **D. Molins-Delgado**, S. Rodríguez-Mozaz, E. Eljarrat, M.S. Díaz-Cruz, D. Barceló, G. Caminal, T. Vicent, "Re-inoculation strategies enhance the degradation of emerging pollutants in fungal bioaugmentation of sewage sludge," *Bioresource Technology*, vol. 168, pp.180-189, 2014.
2. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Removal of polar UV Stabilizers in biological wastewater treatments and ecotoxicological implications," *Chemosphere*, vol. 119, pp. S51-S57, 2015.
3. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Ecological risk assessment associated to the removal of endocrine-disrupting parabens and benzophenone-4 in wastewater treatment," *Journal of Hazardous Materials*, vol. 310, pp. 143-151, 2016.
4. **D. Molins-Delgado**, P. Gago-Ferrero, M.S. Díaz-Cruz, D. Barceló, "Single and joint ecotoxicity data estimation of organic UV filters and nanomaterials toward selected aquatic organisms. Urban groundwater risk assessment," *Environmental Research*, vol.145, pp.126-134, 2016.
5. J. Valle-Sistac, **D. Molins-Delgado**, M. Díaz, L. Ibáñez, D. Barceló, M.S. Díaz-Cruz, "Determination of parabens and benzophenone-type UV filters in human placenta. First determination of the existence of benzyl paraben and benzophenone-4," *Environment International*, vol. 88, pp. 243-249, 2016.
6. L. Mandaric, E. Diamantini, E. Stella, S. Malluci, J. Valle-Sistac, **D. Molins-Delgado**, A. Bellin, G. Chiogna, B. Majone, M.S. Diaz-Cruz, S. Sabater, D. Barceló, M. Petrovic, "Contamination sources and distribution patterns of pharmaceuticals and personal care products in the Alpine rivers strongly affected by tourism," *Science of the Total Environment*, vol. 590-591, pp. 484-494, 2017.
7. **D. Molins-Delgado**, J. Távora, M.S. Díaz-Cruz, D. Barceló, "UV filters and benzotriazoles in urban aquatic ecosystems: the footprint of daily use products," *Science of the Total Environment*, vol. 601-602, pp. 975 – 986, 2017.
8. A. Mizukawa, **D. Molins-Delgado**; J.C. Rodrigues Azevedo; C.V. Scapulatempo Fernandes; M.S. Díaz-Cruz; D. Barceló, "Sediments as a sink for UV filters: The case study of Upper Iguaçú watershed, Curitiba (Brazil)," *Environmental Science and Pollution Research*, vol. 24, pp. 18284-18294, 2017.

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9. **D. Molins-Delgado**, M. Máñez, A. Andreu, F. Hiraldo, E. Eljarrat, M. S. Díaz-Cruz, D. Barceló, "New threat to wild life: UV filters in bird eggs from a preserved area," *Environmental Science and Technology (en prensa)* DOI: 10.1021/acs.est.7b03300.
 10. **D. Molins-Delgado**, D. García-Sillero, M.S. Díaz-Cruz, D. Barceló, "Development of a new on-line SPE-HPLC-(APPI)-MS/MS method for the determination of insect repellents in European rivers," *Journal of Chromatography A* (enviado).
 11. **D. Molins-Delgado**, R. Muñoz, S. Nogueira, M.B. Alonso, J.P. Torres, O. Malm, R.L. Zioli, R.A. Hauser-Davis, E. Eljarrat, D. Barceló, M.S. Díaz-Cruz, "Occurrence of organic UV filters and metabolites in lebranche mullet (*Mugil liza*) from Brazil," *Science of the Total Environment* (enviado).
 12. **D. Molins-Delgado**, M.M. Olmo-Campos, G. Valeta-Juan, V. Pleguezuelos-Hernández, D. Barceló, M.S. Díaz-Cruz, "Determination of UV filters in human breast milk using turbulent flow chromatography and babies' daily intake estimation," *Environmental Research* (enviado).
 13. A.C. Soler de la Vega, **D. Molins-Delgado**, D. Barceló, M.S. Díaz Cruz, "Single and mixture toxicity of organic and inorganic UV-Filters and parabens on *Daphnia magna* and *Phaeodactylum tricornutum*," (en preparación).
 14. Y. Valcárcel, M. López de Alda, S. González-Alonso, A. Silva, J.J. Durán, J. López, **D. Molins-Delgado**, M.S. Díaz-Cruz, J. Sanchís, M. Farré, Ò. Aznar-Aleman, E. Eljarrat, D. Barceló, L. Moreno, "Occurrence of anthropogenic organic contaminants (sunscreens filters, perfluorinated compounds and pirethroids) in inland surface waters from the Northern Antarctic peninsular region," (en preparación).

Capítulos de Libro

1. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Introduction: personal care products in the aquatic environment," In Personal Care Products in the Aquatic Environment, M.S. Díaz-Cruz, D. Barceló (Eds.), *The Handbook of Environmental Chemistry*, Springer International Publishing, pp. 1-34, 2015.
2. M.S. Díaz-Cruz, **D. Molins-Delgado**, "Toxicity and Risk Assessment of Organic UV Filters in Aquatic Ecosystems," In Insights in Ecotoxicology, R.A. Hauser-Davis, T. Parente (Eds.), Science Publishers (CRC Press), (aceptado).

Anexo II. Listado de participación en congresos

Presentaciones orales

(El ponente aparece subrayado)

1. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "HPLC-MS/MS Trace determination of multiclass UV filters and UV blockers in milk," *XII Reunión Científica de la Sociedad Española de Cromatografía y Técnicas Afines (SECyTA)*, Tarragona, España, 2012.
2. **D. Molins-Delgado**, P. Gago-Ferrero, M.S. Díaz-Cruz, D. Barceló, "Determination of toxicity data of ultraviolet filters towards selected aquatic organisms for a preliminary environmental risk assessment," *SETAC Europe 24th Annual Meeting*, Basilea, Suiza, 2014.
3. M.S. Díaz-Cruz, **D. Molins-Delgado**, E. Pastoret, D. Barceló, "Removal of Parabens and Benzophenone-4 from wastewater and environmental impact assessment," *SETAC Europe 24th Annual Meeting*, Basilea, Suiza, 2014.
4. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Analytical strategies based on off-line and on-line (Turboflow[®]) extraction of chromatography-tandem mass spectrometry for the trace determination of UV blockers in milk," *10th Annual LC/MS/MS Workshop on Environmental Applications and Food Safety*, Barcelona, España, 2014.
5. D. Barceló, P. Verlicchi, M. Petrovic, P. Gago-Ferrero, **D. Molins-Delgado**, M.S. Díaz-Cruz, N. Mastroianni, M. Köck-Schulmeyer, C. Postigo, M. López de Alda, S. Pérez, A. Ginebreda, "Fate and risk of pesticides, pharmaceuticals, illicit drugs and personal care products in the Iberian river basins of Ebro and Llobregat: challenges and solutions using advanced treatment technologies," *248th ACS National Meeting*, San Francisco, California, Estados Unidos de América, 2014.
6. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Occurrence and fate of personal care products in the environment," *1st Meeting of Young Researchers from the IDAEA-CSIC*, Barcelona, España, 2015.
7. **D. Molins-Delgado**, F.M. Flores, M.S. Díaz-Cruz, D. Barceló, "UV filters bioaccumulation. The need for metabolites inclusion when carrying out ERA," *SETAC Europe 26th Annual Meeting*, Nantes, Francia, 2016.

Presentaciones en formato póster

1. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Removal of polar UV stabilizers in biological wastewater treatments," *SETAC Europe 23th Annual Meeting*, Glasgow, Reino Unido, 2013.
2. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Removal of personal care products in biological wastewater treatments," *VI Reunión de la Sociedad Española de Espectrometría de Masas (SEEM)*, Úbeda, España, 2013.
3. J. Valle-Sistac, **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Analysis of personal care products in human placental tissue," *SETAC Europe 24th Annual Meeting*, Basilea, Suiza, 2014.
4. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Ecotoxicity and mixture of ultraviolet filters towards selected aquatic organisms," *SETAC Europe 25th Annual Meeting*, Barcelona, España, 2015.
5. **D. Molins-Delgado**, J. Távara, M.S. Díaz-Cruz, D. Barceló, "Occurrence and environmental impact of organic UV filters in urban aquatic ecosystems," *SETAC Europe 25th Annual Meeting*, Barcelona, España, 2015.
6. **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Analysing personal care products in biota by liquid chromatography-atmospheric pressure photoionisation-mass spectrometry," *SETAC Europe 25th Annual Meeting*, Barcelona, España, 2015.
7. **D. Molins-Delgado**, J. Bazzan Arsand, M.S. Díaz-Cruz, L. Jank, M. Martins, R. Hoff, F. Barreto, C. Sirtori, T. Pizzolato, D. Barceló, "UV Aminoglycosides antibiotics residues analysis in bovine milk and bovine, swine and poultry muscle by LC-MS/MS and LC-QTOF-MS: a simple and fast non SPE method," *SETAC Europe 26th Annual Meeting*, Nantes, Francia, 2016.
8. M.P. Serra-Roig, **D. Molins-Delgado**, M.S. Díaz-Cruz, D. Barceló, "Automated LC-MS² analysis of sunscreen residues in salty waters," *12th Annual LC/MS/MS Workshop on Environmental Applications and Food Safety*, Barcelona, España, 2016.

Anexo III. Otras publicaciones (no incluidas en el cuerpo de la Memoria de Tesis)

Publicación Nº A1

Introduction: Personal Care Products in the Aquatic Environment

Por

D. Molins-Delgado, M.S. Díaz-Cruz, D. Barceló

en

Personal Care Products in the Aquatic Environment

M.S. Díaz-Cruz and D. Barceló (Eds.)

The Handbook of Environmental Chemistry

Springer International Publishing

pp. 1-34, 2015

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<http://www.springer.com/gp/book/9783319188089>

Publicación Nº A2

Contamination sources and distribution patterns of pharmaceuticals and personal care products in Alpine rivers strongly affected by tourism

Por

L. Mandaric, E. Diamantini, E. Stella, S. Malluci, J. Valle-Sistac, D. Molins-Delgado, A. Bellin, G. Chiogna, B. Majone, M.S. Diaz-Cruz, S. Sabater, D. Barceló, M. Petrovic

en

Science of the Total Environment, vol. 590-591, pp. 484-494, 2017.

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Anexo IV. Información adicional de las publicaciones

Información adicional - publicación Nº 1

*UV filters and benzotriazoles in urban aquatic ecosystems:
the footprint of daily use products*

por

D. Molins-Delgado, J. Távora, M.S. Díaz-Cruz, D. Barceló

en

Science of the Total Environment, vol. 601-602, pp. 975-986, 2017

Supporting Information
For
UV FILTERS AND BENZOTRIAZOLES IN URBAN AQUATIC
ECOSYSTEMS:
THE FOOTPRINT OF DAILY USE PRODUCTS

Daniel Molins-Delgado¹, João Távora¹, M. Silvia Díaz-Cruz^{1*}, Damià Barceló^{1,2}

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Table A6. Estimated daily mass loads ($\text{mg d}^{-1} \text{inh}^{-1}$) for each compound and the cumulative loads (Σ UV-Fs), WWTP, and population served. n.a.: not applicable.

Table A7. Acute (a) and chronic (b) hazard quotients (HQs) for river waters. -: not available. BP3 (1) and (2) indicate HQs for two different LOEC values.

Table A8. Acute (a) and chronic (b) toxicity hazard quotients for influent (Inf) and effluent (Eff) wastewaters. -: not available. BP3 (1) and (2) indicate HQs for two different LOEC values.

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Table A1. Name of the selected compounds, abbreviation, CAS number, and the log of the partition coefficient octanol water (Log K_{ow}).

Compound	Abbreviation	CAS no.	Log K_{ow}
Benzophenone-1	BP1	131-56-6	3.17
Benzophenone-3	BP3	131-57-7	3.79
4-hydroxybenzophenone	4HB	1137-42-4	3.02
4,4 -dihydroxybenzophenone	4DHB	611-99-4	2.55
4-methylbenzylidene camphor	4MBC	36861-47-9	4.95
Ethylhexyl methoxycinnamate	EHMC	5466-77-3	5.80
Ethylhexyldimethyl PABA	ODPABA	21245-02-3	6.15
Octocrylene	OC	6197-30-4	7.53
Ethyl 4-aminobenzoate	EtPABA	94-09-7	1.86
1H-benzotriazole	BZT	95-14-7	1.23
5-methyl-1-H-benzotriazole	MeBZT	136-85-6	1.89

Table A2. Operational parameters of the studied WWTPs. HRT: hydraulic retention time; -: not available, d: days.

WWTP	Location	Treatment	HRT (d)	Designed Treatment Capacity (m ³ d ⁻¹)	Average Flow (m ³ d ⁻¹) (operated)	Population served	Equivalent inhabitants (design)	Equivalent inhabitants (operated)
S3	Montcada i Reixac	Biologic	0.3	72600	50000	253364	423500	229000
S6	Sabadell	Biologic with P + N removal	-	35000	25500	118675	296333	-
S8	Terrassa	Biologic	29.9	60000	38000	195160	500000	165000
S11	St. Feliu de Llobregat	Biologic with P + N removal	0.36	64000	43000	279959	373333	198000
S14	Rubí	Biologic with N removal	0.3	27000	23000	77994	135000	142000
S17	Manresa	Biologic with P + N removal	-	53500	28000	85224	196.167	-

Table A3. Description of the sampling sites.

Sample name	Type of water sample	Sediment	Location	River	Coordinates
S2	river	Yes	Santa Coloma de Gramanet	Besòs	41.455497 N, 2.194237 E
S3I	influent WWTP	No	Montcada i Reixac	Besòs	41.472330 N, 2.190766 E
S3E	effluent WWTP	No	WWTP		
S4	river	Yes	Montcada i Reixac	Ripoll	41.488159 N, 2.187565 E
S5	river	Yes	Montcada i Reixac	Besòs	41.489726 N, 2.192799 E
S6I	influent WWTP	No	Sabadell	Riu-Sec	41.517075 N, 2.101912 E
S6E	effluent WWTP	No	WWTP		
S7	river	Yes	Sabadell	Riu-Sec	41.51441 N, 2.108685 E
S8I	influent WWTP	No	Terrassa	Riera de Rubí	41.517907 N, 2.034551 E
S8E	effluent WWTP	No	WWTP		
S9	river	Yes	Les Fonts	Riera de Rubí	41.52172 N, 2.037274 E
S10	river	Yes	Les Fonts	Riera de Rubí	41.51161 N, 2.033814 E
S11I	influent WWTP	No	St. Feliu de Llobregat	Llobregat	41.381574 N, 2.033401 E
S11E	effluent WWTP	No	WWTP		
S12	river	Yes	St. Feliu de Llobregat	Llobregat	41.381574 N, 2.033401 E
S13	river	Yes	St. Feliu de Llobregat	Llobregat	41.384444 N, 2.025893 E
S14I	influent WWTP	No	Rubí WWTP	Riera de Rubí	~41.461447 N, ~2.003419 E
S14E	effluent WWTP	No			
S15	river	Yes	Rubí	Riera de Rubí	41.456371 N, 2.001271 E
S16	river	Yes	Rubí	Riera de Rubí	41.460988 N, 2.000863 E
S17I	influent WWTP	No	Manresa	Cardener	41.703772 N, 1.843232 E
S17E	effluent WWTP	No	WWTP		
S18	river	Yes	Manresa	Cardener	41.720803 N, 1.827565 E
S19	river	Yes	Castellgalí	Cardener	41.681077 N, 1.848878 E

Table A4. Instrumental performance for the water (a) and sediment and suspended particulate matter (b) analytical methods.

a)

HPLC-ESI-MS/MS analysis of water samples						
Compound	Linearity range (ng l ⁻¹)	r ²	ILOD (pg)	ILOQ (pg)	Precision (RSD%), n=7	
					Intra-day	Inter-day
BP1	0.5-500	0.9998	10	33	3	6
BP3	0.5-500	0.9998	4	13	3	5
4HB	0.5-500	0.9999	6	20	4	5
4DHB	0.5-500	0.9997	14	47	3	5
DHMB	0.5-500	0.9995	8	27	4	6
4MBC	1-500	0.9995	6	20	3	6
EtPABA	0.1-200	0.9991	0.2	0.7	5	7
BZT	1-10000	0.9995	1.3	4.2	5	6
MeBZT	1-10000	0.9995	0.7	2.3	3	5

ILOD - instrumental limit of detection; ILOQ - instrumental limit of quantification; RSD - relative standard deviation

b)

HPLC-ESI-MS/MS analysis of sediment and suspended particulate matter samples						
<i>Instrumental</i>						
Compound	Linearity range (ng l ⁻¹)	r ²	ILOD (pg)	ILOQ (pg)	Precision (RSD%), n=7	
					Intra-day	Inter-day
BP1	0.5-500	0.9998	10.0	33.0	3	6
BP3	0.5-500	0.9998	4.0	13.0	3	5
4HB	0.5-500	0.9999	6.0	20.0	4	5
4DHB	0.5-500	0.9997	14.0	47.0	3	5
4MBC	1-500	0.9995	6.0	20.0	3	6
OC	1-300	0.998	1.1	3.7	5	7
EHMC	1-500	0.993	1.5	4.9	5	7
OD-PABA	1-100	0.995	4.1	13.5	4	10
EiPABA	0.1-200	0.9991	0.2	0.7	5	7
BZT	1-10000	0.9995	1.3	4.2	5	6
MeBZT	1-10000	0.9995	0.7	2.3	3	5

ILOD - instrumental limit of detection; ILOQ - instrumental limit of quantification; RSD - relative standard deviation

Table A5. Concentrations of UV-Fs and benzotriazole in water samples. <LOQ: below the limit of quantification; n.d.: not detected.

Sample Name	Compound									
	BP1	BP3	4HB	4DHB	4MBC	EtPABA	BZT	MeBZT		
<i>River</i>										
S2	48,2	52,2	12,1	< LOQ	13,1	27,2	2498	4209		
S4	14,5	15,9	< LOQ	< LOQ	13,9	7,7	4270,8	4315,9		
S5	31,2	20,9	10,1	< LOQ	< LOQ	38,6	2855,6	5004		
S7	51,8	43,7	< LOQ	< LOQ	18,2	51,7	4852,8	3844,5		
S9	15,6	15,9	10,5	< LOQ	25,8	6,3	244,6	797,9		
S10	< LOQ	< LOQ	< LOQ	< LOQ	18,8	< LOQ	23,7	199,3		
S12	5,3	< LOQ	< LOQ	< LOQ	< LOQ	n.d.	267,5	721,3		
S13	< LOQ	< LOQ	< LOQ	< LOQ	34,3	< LOQ	198,7	538,8		
S15	24,8	30,1	< LOQ	9,2	13,9	111,9	8529,8	7181,4		
S16	28,3	38,5	< LOQ	< LOQ	< LOQ	44,2	1192,5	1934,8		
S18	< LOQ	4,4	< LOQ	< LOQ	< LOQ	5,5	25	66,9		
S19	< LOQ	5	< LOQ	< LOQ	n.d.	n.d.	262	284		
<i>WWTP</i>										
S3	409,7	515,7	n.d.	n.d.	n.d.	184,2	2493,4	4593,2		
Reixac	211,2	217,8	< LOQ	< LOQ	22	35,8	1469,2	4411,7		
S6	687,9	326,9	10,7	< LOQ	n.d.	149,1	9481,6	6366,2		
Sabadell	23,6	29,7	< LOQ	< LOQ	< LOQ	32,1	2891,5	2376,9		
S8	332	387,1	< LOQ	n.d.	30,1	224,5	2953,7	4342,4		
Terrassa	28	46,1	< LOQ	< LOQ	58,1	66	1172,1	2173,6		
S11	261,1	201	n.d.	n.d.	n.d.	12,3	1495,2	3715,1		
St. Feliu de Llobregat	10,2	33,6	< LOQ	< LOQ	< LOQ	23,4	1084,1	4832,8		
S14	94,6	75,5	n.d.	7,7	n.d.	53,3	8281,9	5143,7		
Rubi	5,2	9,4	< LOQ	10,9	34,6	129,1	16933,1	8913,3		
S17	379,7	297,1	24,8	n.d.	29,3	114,8	3272,6	3728,5		
Manresa	4,3	27,1	< LOQ	< LOQ	n.d.	22,4	5100,4	2385,9		

Table A6. Estimated daily mass loads (mg d⁻¹ inh⁻¹) for each compound and the cumulative loads (ΣUV-Fs), WWTP, and population served. n.a.: not applicable.

		Mass Loads (mg d ⁻¹ inh ⁻¹)									
WWTP		BP1	BP3	4HB	4DHB	4MBC	EPABA	BZT	MeBZT	ΣUV-Fs	
S3	Montcada i	0.008	0.010	n.a.	n.a.	n.a.	0.004	0.05	0.09	0.16	
	Reixac	0.004	0.004	n.a.	n.a.	0.0004	0.0007	0.03	0.09	0.13	
S6	inf.	0.010	0.007	0.0002	n.a.	n.a.	0.003	0.20	0.13	0.36	
	Sabadell	0.0005	0.0006	n.a.	n.a.	n.a.	0.0007	0.06	0.05	0.11	
S8	inf.	0.006	0.008	n.a.	n.a.	0.0006	0.004	0.06	0.08	0.16	
	Terrassa	0.0005	0.0009	n.a.	n.a.	0.001	0.001	0.02	0.04	0.07	
S11	St. Feliu de	0.004	0.003	n.a.	n.a.	n.a.	0.0002	0.02	0.06	0.09	
	Llobregat	0.0002	0.0005	n.a.	n.a.	n.a.	0.0004	0.02	0.07	0.09	
S14	inf.	0.0030	0.002	n.a.	0.0002	n.a.	0.002	0.24	0.15	0.40	
	Rubí	0.0002	0.0003	n.a.	0.0003	0.001	0.004	0.50	0.26	0.77	
S17	inf.	0.010	0.010	0.0008	n.a.	0.001	0.004	0.11	0.12	0.26	
	Manresa	0.0001	0.0009	n.a.	n.a.	n.a.	0.0007	0.17	0.08	0.25	

Table A7. Acute (a) and chronic (b) hazard quotients (HQs) for river waters. -: not available. BP3 (1) and (2) indicate HQs for two different LOEC values.

a)

Sample	LC ₅₀ (mg l ⁻¹)	<i>Daphnia magna</i>			<i>Daphnia galeata</i>		<i>Vibrio fischeri</i>	
		BP3	4MBC	BZT	MeBZT	BZT	MeBZT	MeBZT
S2	Besòs	0.03	0.02	0.02	0.08	0.16	0.49	0.48
S4	Ripoll	0.01	0.02	0.04	0.08	0.27	0.50	0.50
S5	Besòs	0.01	-	0.03	0.10	0.18	0.58	0.58
S7	Riu-Sec	0.02	0.03	0.05	0.07	0.31	0.45	0.44
S9	Riera de Rubí	0.01	0.05	2E-03	0.02	0.02	0.09	0.09
S10	Riera de Rubí	-	0.03	2E-04	4E-03	2E-03	0.02	0.02
S12	Llobregat	-	-	3E-03	0.01	0.02	0.08	0.08
S13	Llobregat	-	0.06	2E-03	0.01	0.01	0.06	0.06
S15	Riera de Rubí	0.02	0.02	0.08	0.14	0.54	0.84	0.83
S16	Riera de Rubí	0.02	-	0.01	0.04	0.08	0.23	0.22
S18	Cardener	2E-03	-	2E-04	1E-03	2E-03	0.01	0.01
S19	Cardener	3E-03	-	2E-03	0.01	0.02	0.03	0.03

b)

Sample	<i>Pimephales promelas</i>			<i>Oncorhynchus mykiss</i>				
	BP1	ETPABA	4394	BP1	BP3 (1)	BP3 (2)	4MBC	
	4919.9	4919.9	4394	4919	3900	749	415	
S2	Besòs	0.01	0.01	0.01	0.01	0.07	0.03	
S4	Ripoll	3E-03	2E-03	3E-03	4E-03	0.02	0.03	
S5	Besòs	0.01	0.01	0.01	0.01	0.03	-	
S7	Riu-Sec	0.01	0.01	0.01	0.01	0.06	0.04	
S9	Riera de Rubí	3E-03	-	3E-03	4E-03	0.02	0.06	
S10	Riera de Rubí	-	-	-	-	-	0.05	
S12	Llobregat	1E-03	-	1E-03	-	-	-	
S13	Llobregat	-	3E-02	-	-	-	0.08	
S15	Riera de Rubí	0.01	0.03	0.01	0.01	0.04	0.03	
S16	Riera de Rubí	0.01	0.01	0.01	0.01	0.05	-	
S18	Cardener	-	1E-03	-	-	0.01	-	
S19	Cardener	-	-	-	-	0.01	-	

Table A8. Acute (a) and chronic (b) toxicity hazard quotients for influent (Inf) and effluent (Eff) wastewaters. -: not available. BP3 (1) and (2) indicate HQs for two different LOEC values.

a)

Sample	LC ₅₀ (mg l ⁻¹)	<i>Daphnia magna</i>						<i>Daphnia galeata</i>						<i>Vibrio fischeri</i>	
		BP3		4MBC		BZT		MeBZT		BZT		MeBZT		MeBZT	
		Inf	Eff	Inf	Eff	Inf	Eff	Inf	Eff	Inf	Eff	Inf	Eff	Inf	Eff
S3	Montcada i Reixac	0.27	0.11	-	0.04	0.02	0.01	0.09	0.09	0.16	0.09	0.54	0.51	0.53	0.51
S6	Sabadell	0.17	0.02	-	-	0.09	0.03	0.12	0.05	0.60	0.18	0.74	0.28	0.73	0.27
S8	Terrassa	0.20	0.02	0.05	0.10	0.03	0.01	0.08	0.04	0.19	0.07	0.51	0.25	0.50	0.25
S11	St. Feliu de Llobregat	0.11	0.02	-	-	0.01	0.01	0.07	0.09	0.09	0.07	0.43	0.56	0.43	0.56
S14	Rubi	0.04	5E-03	-	0.06	0.08	0.16	0.10	0.17	0.52	1.07	0.60	1.04	0.59	1.02
S17	Manresa	0.16	0.01	0.05	-	0.03	0.05	0.07	0.05	0.21	0.32	0.43	0.28	0.43	0.27

b)

Sample	<i>Pimephales promelas</i>						<i>Oncorhynchus mykiss</i>							
	BP1		EtPABA		BP3 (1)		BP3 (2)		BP1		4MBC			
	Inf	Eff	Inf	Eff	Inf	Eff	Inf	Eff	Inf	Eff	Inf	Eff		
S3	0.08	0.04	0.04	0.01	0.08	0.04	0.13	0.06	0.08	0.04	0.69	0.29	-	0.05
S6	0.14	5E-03	0.03	0.01	0.14	5E-03	0.08	0.01	0.14	5E-03	0.44	0.04	-	-
S8	0.07	0.01	0.05	0.02	0.07	0.01	0.10	0.01	0.07	0.01	0.52	0.06	0.07	0.14
S11	0.05	2E-03	3E-03	0.01	0.05	2E-03	0.05	0.01	0.05	2E-03	0.27	0.04	-	-
S14	0.02	1E-03	0.01	0.03	0.02	1E-03	0.02	2E-03	0.02	1E-03	0.10	0.01	-	0.08
S17	0.08	9E-04	0.03	0.01	0.08	9E-04	0.08	0.01	0.08	9E-04	0.40	0.04	0.07	-

Información adicional - publicación Nº 2

Development of a new on-line SPE-HPLC-(APPI)-MS/MS method for the determination of insect repellents in European rivers

Por

D. Molins-Delgado, D. García-Sillero, M.S. Díaz-Cruz, D. Barceló

en

Journal of Chromatography A (enviado)

Supporting Information

**A SPE-HPLC-(APPI)-MS/MS method for the analysis of
selected anthropogenic infochemicals in water**

Daniel Molins-Delgado¹, Daniel García-Sillero¹, M. Silvia Díaz-Cruz^{1*}, Damià Barceló^{1,2}

Content

Table A1. Sample name, sampling description. n.a.: not available.

Table A2. IRs concentrations (ng l^{-1}) in the water samples.

Table A3. Estimated hazard quotients (acute and chronic toxicity). EC₅₀: half maximal effective concentration; LC₅₀: half maximal lethal concentration; NOEC: No-observed-adverse-effects; n.a.: not applicable; n.s.: not specified.

References

Table A1. Sample name, sampling description. n.a.: not available.

River Basin	Sample	Coordinates		Description
		Latitude	Longitude	
Sava	RAD2	46.29298	14.260754	Background location
	LIT1	46.08456	15.582632	Land use
	CAT1	45.89636	15.627037	Land use
	CAT2	45.86036	15.692068	
	ZAG1	45.78301	16.003184	Urban centre
	CRN1	n.a.	n.a.	Navigation; oil refinery; land use
	SLB1	45.15270	18.013906	Navigation
	SLB2	45.12622	18.084751	
	ZUP1	45.07481	18.687667	Urban centre; navigation; oil refinery
	ZUP2	45.01488	18.740313	
	SRM2	44.91358	19.752412	Navigation
	BEO1	44.76913	20.355572	Urban centre; navigation
BEO2	44.80632	20.443683		
Evrotas	USkollini7	n.a.	n.a.	Reference drought location; woodlands
	USkollino22	n.a.	n.a.	
	DSkollini9	n.a.	n.a.	Land use
	DSkallio29	n.a.	n.a.	
	Vivariie10	n.a.	n.a.	Reference pollution location; land use
	Vivarioe15	n.a.	n.a.	
	WWTPi19	n.a.	n.a.	Urban centre; land use WWTP
WWTPo5	n.a.	n.a.		
Adige	wb2a	46.25872	10.604122	High course of the river; holidays resorts
	wb2b/w1	46.25971	10.608057	
	wb3a	46.31971	10.818326	
	wb3b-1	46.32844	10.870947	
	wb1	46.42556	10.928332	Land use; holidays resorts
	wb4b	46.21917	11.101943	
	wb5b	46.15722	11.076386	
	wb6b	46.19250	11.128336	
	wb7a	46.00960	11.122896	
	wb7a-1	45.94550	11.101080	
wb7a-x	45.91262	11.038397	Urban centre	
wb7b-1	45.88857	11.015244		

Table A2. IRs concentrations (ng l⁻¹) in the water samples.

River Basin	Sample	Concentrations (ng l ⁻¹)				
		DEET	m-toluamide	Bayrepel	MGK-264	PBO
Sava	RAD2	70.80	n.d.	103.80	n.d.	n.d.
	LIT1	39.23	n.d.	72.23	n.d.	n.d.
	CAT1	12.25	n.d.	31.77	n.d.	n.d.
	CAT2	8.92	n.d.	18.22	n.d.	n.d.
	ZAG1	57.37	n.d.	57.28	n.d.	n.d.
	CRN1	61.49	n.d.	105,336.01	n.d.	n.d.
	SLB1	12.34	n.d.	9.26	n.d.	n.d.
	SLB2	1.18	n.d.	2.23	n.d.	n.d.
	ZUP1	26.65	n.d.	266.31	n.d.	n.d.
	ZUP2	3.44	n.d.	148.48	n.d.	n.d.
	SRM2	7.69	n.d.	28.75	n.d.	n.d.
	BEO1	53.72	n.d.	314.69	n.d.	n.d.
	BEO2	8.33	n.d.	18.86	n.d.	n.d.
	Evrotas	USkollini7	2,005.45	n.d.	17.36	n.d.
USkollino22		59.22	n.d.	31.93	n.d.	n.d.
DSkollini9		398.98	n.d.	179.62	n.d.	n.d.
DSkallio29		4,348.82	n.d.	44.35	n.d.	n.d.
Vivariie10		636.37	n.d.	15.70	n.d.	n.d.
Vivarioe15		510.10	n.d.	21.40	n.d.	19.33
WWTPi19		414.09	n.d.	13.23	n.d.	n.d.
WWTPo5		4,948.85	n.d.	6.48	n.d.	n.d.
Adige	wb2a	n.d.	n.d.	n.d.	n.d.	n.d.
	wb2b/w1	6.80	n.d.	2.92	n.d.	n.d.
	wb3a	n.d.	n.d.	n.d.	n.d.	n.d.
	wb3b-1	n.d.	n.d.	n.d.	n.d.	n.d.
	wb1	n.d.	n.d.	n.d.	n.d.	n.d.
	wb4b	n.d.	n.d.	n.d.	n.d.	n.d.
	wb5b	n.d.	n.d.	37.38	n.d.	n.d.
	wb6b	n.d.	n.d.	n.d.	n.d.	n.d.
	wb7a	n.d.	n.d.	n.d.	n.d.	n.d.
	wb7a-1	n.d.	n.d.	n.d.	n.d.	n.d.
	wb7a-x	6.01	n.d.	n.d.	n.d.	n.d.
wb7b-1	n.d.	n.d.	n.d.	n.d.	n.d.	

Table A3. Estimated hazard quotients (acute and chronic toxicity). EC₅₀: half maximal effective concentration; LC₅₀: half maximal lethal concentration; NOEC: No-observed-adverse-effects; n.a.: not applicable; n.s.: not specified.

		HQ				
Acute Toxicity Data	Species	Daphnia magna	Daphnia magna	Daphnia magna	Pseudokirchneriella subcapitata	Vibrio fischeri
	EC₅₀/LC₅₀ (mg l⁻¹)	75	34.4	108	4.1	21.2
	Criteria	Intoxication	Intoxication	48 h mortality	Growth Inhibition	Disminution of luminiscence
	Reference	[1]	[2]	[3]	[4]	[4]
Basin	Sample	DEET	DEET	DEET	DEET	DEET
Sava	RAD2	9.4E-04	2.1E-03	6.6E-04	1.7E-02	3.3E-03
	LIT1	5.2E-04	1.1E-03	3.6E-04	9.6E-03	1.9E-03
	CAT1	1.6E-04	3.6E-04	1.1E-04	3.0E-03	5.8E-04
	CAT2	1.2E-04	2.6E-04	8.3E-05	2.2E-03	4.2E-04
	ZAG1	7.6E-04	1.7E-03	5.3E-04	1.4E-02	2.7E-03
	CRN1	8.2E-04	1.8E-03	5.7E-04	1.5E-02	2.9E-03
	SLB1	1.6E-04	3.6E-04	1.1E-04	3.0E-03	5.8E-04
	SLB2	1.6E-05	3.4E-05	1.1E-05	2.9E-04	5.6E-05
	ZUP1	3.6E-04	7.7E-04	2.5E-04	6.5E-03	1.3E-03
	ZUP2	4.6E-05	1.0E-04	3.2E-05	8.4E-04	1.6E-04
	SRM2	1.0E-04	2.2E-04	7.1E-05	1.9E-03	3.6E-04
	BEO1	7.2E-04	1.6E-03	5.0E-04	1.3E-02	2.5E-03
BEO2	1.1E-04	2.4E-04	7.7E-05	2.0E-03	3.9E-04	
Evrotas	USkollini7	2.7E-02	5.8E-02	1.9E-02	0.49	9.5E-02
	USkollino22	7.9E-04	1.7E-03	5.5E-04	1.4E-02	2.8E-03
	DSkollini9	5.3E-03	1.2E-02	3.7E-03	9.7E-02	1.9E-02
	DSkallio29	5.8E-02	0.13	4.0E-02	1.1	0.21
	Vivariie10	8.5E-03	1.8E-02	5.9E-03	0.16	3.0E-02
	Vivarioe15	6.8E-03	1.5E-02	4.7E-03	0.12	2.4E-02
	WWTPi19	5.5E-03	1.2E-02	3.8E-03	0.1	2.0E-02
	WWTPo5	6.6E-02	0.14	4.6E-02	1.2	0.23
Adige	wb2a	n.a.	n.a.	n.a.	n.a.	n.a.
	wb2b/w1	9.1E-05	2.0E-04	6.3E-05	1.7E-03	3.2E-04
	wb3a	n.a.	n.a.	n.a.	n.a.	n.a.
	wb3b-1	n.a.	n.a.	n.a.	n.a.	n.a.
	wb1	n.a.	n.a.	n.a.	n.a.	n.a.
	wb4b	n.a.	n.a.	n.a.	n.a.	n.a.
	wb5b	n.a.	n.a.	n.a.	n.a.	n.a.
	wb6b	n.a.	n.a.	n.a.	n.a.	n.a.
	wb7a	n.a.	n.a.	n.a.	n.a.	n.a.
	wb7a-1	n.a.	n.a.	n.a.	n.a.	n.a.
	wb7a-x	8.0E-05	1.7E-04	5.6E-05	1.5E-03	2.8E-04
wb7b-1	n.a.	n.a.	n.a.	n.a.	n.a.	

Table A3. (Continued).

		HQ				
Acute Toxicity Data	Species	Photobacterium phosphoreum	Onchorhynchus mykiss	Pimephales promelas	Gambusia affinis	Oreochromis mossambicus
	EC₅₀/LC₅₀ (mg l⁻¹)	67.9	71.25	75.7	235	120
	Criteria	Disminution of luminiscence	96 h mortality	96 h mortality	48 h mortality	96 h static renewal
	Reference	[5]	[1]	[6]	[7]	[8]
Basin	Sample	DEET	DEET	DEET	DEET	DEET
Sava	RAD2	1.0E-03	9.9E-04	9.4E-04	3.0E-04	5.9E-04
	LIT1	5.8E-04	5.5E-04	5.2E-04	1.7E-04	3.3E-04
	CAT1	1.8E-04	1.7E-04	1.6E-04	5.2E-05	1.0E-04
	CAT2	1.3E-04	1.3E-04	1.2E-04	3.8E-05	7.4E-05
	ZAG1	8.4E-04	8.1E-04	7.6E-04	2.4E-04	4.8E-04
	CRN1	9.1E-04	8.6E-04	8.1E-04	2.6E-04	5.1E-04
	SLB1	1.8E-04	1.7E-04	1.6E-04	5.3E-05	1.0E-04
	SLB2	1.7E-05	1.7E-05	1.6E-05	5.0E-06	9.8E-06
	ZUP1	3.9E-04	3.7E-04	3.5E-04	1.1E-04	2.2E-04
	ZUP2	5.1E-05	4.8E-05	4.5E-05	1.5E-05	2.9E-05
	SRM2	1.1E-04	1.1E-04	1.0E-04	3.3E-05	6.4E-05
	BEO1	7.9E-04	7.5E-04	7.1E-04	2.3E-04	4.5E-04
	BEO2	1.2E-04	1.2E-04	1.1E-04	3.5E-05	6.9E-05
Evrotas	USkollini7	3.0E-02	2.8E-02	2.6E-02	8.5E-03	1.7E-02
	USkollino22	8.7E-04	8.3E-04	7.8E-04	2.5E-04	4.9E-04
	DSkollini9	5.9E-03	5.6E-03	5.3E-03	1.7E-03	3.3E-03
	DSkallio29	6.4E-02	6.1E-02	5.7E-02	1.9E-02	3.6E-02
	Vivariie10	9.4E-03	8.9E-03	8.4E-03	2.7E-03	5.3E-03
	Vivarioe15	7.5E-03	7.2E-03	6.7E-03	2.2E-03	4.3E-03
	WWTPi19	6.1E-03	5.8E-03	5.5E-03	1.8E-03	3.5E-03
	WWTPo5	7.3E-02	6.9E-02	6.5E-02	2.1E-02	4.1E-02
Adige	wb2a	n.a.	n.a.	n.a.	n.a.	n.a.
	wb2b/w1	1.0E-04	9.5E-05	9.0E-05	2.9E-05	5.7E-05
	wb3a	n.a.	n.a.	n.a.	n.a.	n.a.
	wb3b-1	n.a.	n.a.	n.a.	n.a.	n.a.
	wb1	n.a.	n.a.	n.a.	n.a.	n.a.
	wb4b	n.a.	n.a.	n.a.	n.a.	n.a.
	wb5b	n.a.	n.a.	n.a.	n.a.	n.a.
	wb6b	n.a.	n.a.	n.a.	n.a.	n.a.
	wb7a	n.a.	n.a.	n.a.	n.a.	n.a.
	wb7a-1	n.a.	n.a.	n.a.	n.a.	n.a.
	wb7a-x	8.9E-05	8.4E-05	7.9E-05	2.6E-05	5.0E-05
	wb7b-1	n.a.	n.a.	n.a.	n.a.	n.a.

Table A3. (Continued).

HQ					
Acute Toxicity Data	Species	Chironomus riparius	Daphnid	Algae	Fish
	EC₅₀/LC₅₀ (mg l⁻¹)	70	34.7	32.13	87.97
	Criteria	96 h	Estimated	Estimated	Estimated
	Reference	[9]	ECOSAR v1.11	ECOSAR v1.11	ECOSAR v1.11
Basin	Sample	DEET	Bayrepel	Bayrepel	Bayrepel
Sava	RAD2	1.0E-03	3.0E-03	3.2E-03	1.2E-03
	LIT1	5.6E-04	2.1E-03	2.2E-03	8.2E-04
	CAT1	1.8E-04	9.2E-04	9.9E-04	3.6E-04
	CAT2	1.3E-04	5.3E-04	5.7E-04	2.1E-04
	ZAG1	8.2E-04	1.7E-03	1.8E-03	6.5E-04
	CRN1	8.8E-04	3.0	3.3	1.2
	SLB1	1.8E-04	2.7E-04	2.9E-04	1.1E-04
	SLB2	1.7E-05	6.4E-05	6.9E-05	2.5E-05
	ZUP1	3.8E-04	7.7E-03	8.3E-03	3.0E-03
	ZUP2	4.9E-05	4.3E-03	4.6E-03	1.7E-03
	SRM2	1.1E-04	8.3E-04	8.9E-04	3.3E-04
	BEO1	7.7E-04	9.1E-03	9.8E-03	3.6E-03
	BEO2	1.2E-04	5.4E-04	5.9E-04	2.1E-04
Evrotas	USkollini7	2.9E-02	5.0E-04	5.4E-04	2.0E-04
	USkollino22	8.5E-04	9.2E-04	9.9E-04	3.6E-04
	DSkollini9	5.7E-03	5.2E-03	5.6E-03	2.0E-03
	DSkallio29	6.2E-02	1.3E-03	1.4E-03	5.0E-04
	Vivariie10	9.1E-03	4.5E-04	4.9E-04	1.8E-04
	Vivarioe15	7.3E-03	6.2E-04	6.7E-04	2.4E-04
	WWTPi19	5.9E-03	3.8E-04	4.1E-04	1.5E-04
	WWTPo5	7.1E-02	1.9E-04	2.0E-04	7.4E-05
Adige	wb2a	n.a.	n.a.	n.a.	n.a.
	wb2b/w1	9.7E-05	8.4E-05	9.1E-05	3.3E-05
	wb3a	n.a.	n.a.	n.a.	n.a.
	wb3b-1	n.a.	n.a.	n.a.	n.a.
	wb1	n.a.	n.a.	n.a.	n.a.
	wb4b	n.a.	n.a.	n.a.	n.a.
	wb5b	n.a.	1.1E-03	1.2E-03	4.2E-04
	wb6b	n.a.	n.a.	n.a.	n.a.
	wb7a	n.a.	n.a.	n.a.	n.a.
	wb7a-1	n.a.	n.a.	n.a.	n.a.
	wb7a-x	8.6E-05	n.a.	n.a.	n.a.
	wb7b-1	n.a.	n.a.	n.a.	n.a.

Table A3. (Continued).

		HQ			
Acute Toxicity Data	Species	Pseudokirchneriella subcapitata	Pseudokirchneriella subcapitata	Pseudokirchneriella subcapitata	Pseudokirchneriella subcapitata
	NOEC (mg l⁻¹)	7.6	3.8	15	0.521
	Criteria	Cell density	Biomass	Growth Inhibition	Growth Inhibition
	Reference	[10]	[10]	[10]	[4]
Basin	Sample	DEET	DEET	DEET	DEET
Sava	RAD2	9.3E-07	1.9E-05	4.7E-06	1.4E-04
	LIT1	5.2E-07	1.0E-05	2.6E-06	7.5E-05
	CAT1	1.6E-07	3.2E-06	8.2E-07	2.4E-05
	CAT2	1.2E-07	2.3E-06	5.9E-07	1.7E-05
	ZAG1	7.5E-07	1.5E-05	3.8E-06	1.1E-04
	CRN1	8.1E-07	1.6E-05	4.1E-06	1.2E-04
	SLB1	1.6E-07	3.2E-06	8.2E-07	2.4E-05
	SLB2	1.6E-08	3.1E-07	7.9E-08	2.3E-06
	ZUP1	3.5E-07	7.0E-06	1.8E-06	5.1E-05
	ZUP2	4.5E-08	9.1E-07	2.3E-07	6.6E-06
	SRM2	1.0E-07	2.0E-06	5.1E-07	1.5E-05
	BEO1	7.1E-07	1.4E-05	3.6E-06	1.0E-04
	BEO2	1.1E-07	2.2E-06	5.6E-07	1.6E-05
	Evrotas	USkollini7	2.6E-05	5.3E-04	1.3E-04
USkollino22		7.8E-07	1.6E-05	3.9E-06	1.1E-04
DSkollini9		5.2E-06	1.0E-04	2.7E-05	7.7E-04
DSkallio29		5.7E-05	1.1E-03	2.9E-04	8.3E-03
Vivariie10		8.4E-06	1.7E-04	4.2E-05	1.2E-03
Vivarioe15		6.7E-06	1.3E-04	3.4E-05	9.8E-04
WWTPi19		5.4E-06	1.1E-04	2.8E-05	7.9E-04
WWTPo5		6.5E-05	1.3E-03	3.3E-04	9.5E-03
Adige	wb2a	n.a.	n.a.	n.a.	n.a.
	wb2b/w1	8.9E-08	1.8E-06	4.5E-07	1.3E-05
	wb3a	n.a.	n.a.	n.a.	n.a.
	wb3b-1	n.a.	n.a.	n.a.	n.a.
	wb1	n.a.	n.a.	n.a.	n.a.
	wb4b	n.a.	n.a.	n.a.	n.a.
	wb5b	n.a.	n.a.	n.a.	n.a.
	wb6b	n.a.	n.a.	n.a.	n.a.
	wb7a	n.a.	n.a.	n.a.	n.a.
	wb7a-1	n.a.	n.a.	n.a.	n.a.
	wb7a-x	7.9E-08	1.6E-06	4.0E-07	1.2E-05
	wb7b-1	n.a.	n.a.	n.a.	n.a.

Table A3. (Continued).

		HQ			
Chronic Toxicity Data	Species	Daphnia magna	Daphnia magna	Algae	Fish
	EC₅₀/LC₅₀ (mg l⁻¹)	26	3.7	9.65	8.42
	Criteria	21 d reproduction	n.s.	Estimated	Estimated
	Reference	[11]	[12]	[12]	[12]
Basin	Sample	DEET	DEET	DEET	DEET
Sava	RAD2	2.7E-03	1.9E-02	7.3E-03	8.4E-03
	LIT1	1.5E-03	1.1E-02	4.1E-03	4.7E-03
	CAT1	4.7E-04	3.3E-03	1.3E-03	1.5E-03
	CAT2	3.4E-04	2.4E-03	9.2E-04	1.1E-03
	ZAG1	2.2E-03	1.6E-02	5.9E-03	6.8E-03
	CRN1	2.4E-03	1.7E-02	6.4E-03	7.3E-03
	SLB1	4.7E-04	3.3E-03	1.3E-03	1.5E-03
	SLB2	4.5E-05	3.2E-04	1.2E-04	1.4E-04
	ZUP1	1.0E-03	7.2E-03	2.8E-03	3.2E-03
	ZUP2	1.3E-04	9.3E-04	3.6E-04	4.1E-04
	SRM2	3.0E-04	2.1E-03	8.0E-04	9.1E-04
	BEO1	2.1E-03	1.5E-02	5.6E-03	6.4E-03
	BEO2	3.2E-04	2.3E-03	8.6E-04	9.9E-04
Evrotas	USkollini7	7.7E-02	0.54	0.21	0.24
	USkollino22	2.3E-03	1.6E-02	6.1E-03	7.0E-03
	DSkollini9	1.5E-02	0.11	4.1E-02	4.7E-02
	DSkallio29	0.17	1.20	0.45	0.52
	Vivariie10	2.4E-02	0.17	6.6E-02	7.6E-02
	Vivarioe15	2.0E-02	0.14	5.3E-02	6.1E-02
	WWTPi19	1.6E-02	0.11	4.3E-02	4.9E-02
WWTPo5	0.19	1.30	0.51	0.59	
Adige	wb2a	n.a.	n.a.	n.a.	n.a.
	wb2b/w1	2.6E-04	1.8E-03	7.0E-04	8.1E-04
	wb3a	n.a.	n.a.	n.a.	n.a.
	wb3b-1	n.a.	n.a.	n.a.	n.a.
	wb1	n.a.	n.a.	n.a.	n.a.
	wb4b	n.a.	n.a.	n.a.	n.a.
	wb5b	n.a.	n.a.	n.a.	n.a.
	wb6b	n.a.	n.a.	n.a.	n.a.
	wb7a	n.a.	n.a.	n.a.	n.a.
	wb7a-1	n.a.	n.a.	n.a.	n.a.
	wb7a-x	2.3E-04	1.6E-03	6.2E-04	7.1E-04
wb7b-1	n.a.	n.a.	n.a.	n.a.	

Table A3. (Continued).

		HQ		
Chronic Toxicity Data	Species	Daphnid	Algae	Fish
	EC ₅₀ /LC ₅₀ (mg l ⁻¹)	3.92	9.46	6.03
	Criteria	Estimated	Estimated	Estimated
	Reference	ECOSAR v1.11	ECOSAR v1.11	ECOSAR v1.11
Basin	Sample	Bayrepel	Bayrepel	Bayrepel
Sava	RAD2	2.6E-02	1.1E-02	1.7E-02
	LIT1	1.8E-02	7.6E-03	1.2E-02
	CAT1	8.1E-03	3.4E-03	5.3E-03
	CAT2	4.6E-03	1.9E-03	3.0E-03
	ZAG1	1.5E-02	6.1E-03	9.5E-03
	CRN1	0.27	0.11	0.17
	SLB1	2.4E-03	9.8E-04	1.5E-03
	SLB2	5.7E-04	2.4E-04	3.7E-04
	ZUP1	6.8E-02	2.8E-02	4.4E-02
	ZUP2	3.8E-02	1.6E-02	2.5E-02
	SRM2	7.3E-03	3.0E-03	4.8E-03
	BEO1	8.0E-02	3.3E-02	5.2E-02
	BEO2	4.8E-03	2.0E-03	3.1E-03
	Evrotas	USkollini7	4.4E-03	1.8E-03
USkollino22		8.1E-03	3.4E-03	5.3E-03
DSkollini9		4.6E-02	1.9E-02	3.0E-02
DSkallio29		1.1E-02	4.7E-03	7.4E-03
Vivariie10		4.0E-03	1.7E-03	2.6E-03
Vivarioe15		5.5E-03	2.3E-03	3.5E-03
WWTPi19		3.4E-03	1.4E-03	2.2E-03
WWTPo5		1.7E-03	6.8E-04	1.1E-03
Adige	wb2a	n.a.	n.a.	n.a.
	wb2b/w1	7.4E-04	3.1E-04	4.8E-04
	wb3a	n.a.	n.a.	n.a.
	wb3b-1	n.a.	n.a.	n.a.
	wb1	n.a.	n.a.	n.a.
	wb4b	n.a.	n.a.	n.a.
	wb5b	9.5E-03	4.0E-03	6.2E-03
	wb6b	n.a.	n.a.	n.a.
	wb7a	n.a.	n.a.	n.a.
	wb7a-1	n.a.	n.a.	n.a.
	wb7a-x	n.a.	n.a.	n.a.
	wb7b-1	n.a.	n.a.	n.a.

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Información adicional - publicación Nº 3

Sediments as a sink for UV filters and benzotriazoles: The case study of Upper Iguaçu watershed, Curitiba (Brazil)

Por

A. Mizukawa, D. Molins-Delgado; J.C. Rodrigues Azevedo; C.V. Scapulatempo Fernandes;
M.S. Díaz-Cruz; D. Barceló

en

Environmental Science and Pollution Research, vol. 24, pp. 18284-18294, 2017

SUPPORTING INFORMATION

Sediments as a sink for UV filters and benzotriazoles: The case study of Upper Iguaçú watershed, Curitiba (Brazil)

Alinne Mizukawa, Daniel Molins-Delgado, Júlio César Rodrigues de Azevedo, Cristóvão Vicente Scapulatempo Fernandes, M. Silvia Díaz-Cruz, Damià Barceló

Table S1: SRM transitions and operational MS/MS parameters for the detection of target UV filters and benzotriazoles. DP (V): Declustering Potential; CE (V): Collision Energy; CXP (V): Transfer potential. In bold, transitions employed for quantitative analysis. Grey cells denote the internal standard compounds used.

Compound	Precursors m/z	Product m/z	DP (V)	CE (V)	CXP (V)
BP1	215	137	40	27	10
	215	105	40	29	6
BP3	229	151	40	25	12
	229	105	40	27	16
4HB	199	121	40	25	8
	199	105	40	27	8
DHMB	245	151	43	27	12
	245	121	43	29	8
4MBC	255	105	61	41	6
	255	212	61	29	14
EHMC	291	179	51	13	4
	291	161	51	25	10
OC	362	250	71	15	20
	362	232	71	57	12
OD-PABA	278	151	86	43	42
	278	136	86	27	40
EtPABA	166	138	41	20	10
	166	120	41	25	28
BZT	120	92	56	25	16
	120	65	56	31	4
MeBZT	134	95	46	39	16
	134	79	46	29	10
TBHPBT	268	212	46	49	4
	268	57	46	49	4
BP3-d ₅	234	151	36	27	12
	234	110	36	27	8
4MBC-d ₄	259	216	76	27	14
	259	109	76	45	4
AllylBzt	266	119	51	31	8
	266	91	51	41	16

Información adicional - publicación N°4

*Removal of polar UV stabilizers in biological wastewater treatments
and ecotoxicological implications*

Por

D. Molins-Delgado, M.S. Díaz-Cruz, D. Barceló

en

Chemosphere, vol. 119, pp. S51-S57, 2015

SUPPORTING INFORMATION

for

REMOVAL OF POLAR UV STABILIZERS IN

BIOLOGICAL WASTEWATER TREATMENTS AND

ECOTOXICOLOGICAL IMPLICATIONS

Daniel Molins-Delgado¹, Silvia Díaz-Cruz^{1*}, Damiá Barceló^{1,2}

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TABLE S2 Concentrations (ngL⁻¹) of BZT and MeBZT in influent and effluent wastewater samples for each studied WWTP.

TABLE S3 Occurrence and removal rates of both BZTs in wastewater of this study and the literature.

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TABLE S1

Table S1a. Instrumental performance of the HPLC-QqLIT-MS/MS analytical method developed. Instrumental limit of detection (ILOD); instrumental limit of quantification (ILOQ); relative standard deviation (RSD).

	linearity (ngL ⁻¹)	r2	ILOD (pg)	ILOQ (pg)	Precision (%RSD) n=5	
					Intraday	Interday
BZT	1-10000	0.999	1.3	4.3	5	6
MeBZT	1-10000	0.999	0.7	2.3	3	5

Table S1b. Recovery rates, method limits of detection (MLOD) and quantification (MLOQ) and matrix effects (ME%) in influent and effluent wastewater samples for BZT and MeBZT, as well as for the IS (AllylBZT).

Concentration (ngL ⁻¹)	BZT			MeBZT			IS ME%			
	RE% ±RSD	MLOD (ngL ⁻¹)	MLOQ (ngL ⁻¹)	RE%±RSD	MLOD (ngL ⁻¹)	MLOQ (ngL ⁻¹)		ME %		
		ME %	ME %							
Influent	5	102±1	1.1	3.7	29	85±7	1.1	3.7	26	21
	50	101±4				89±3				
	100	100±6				91±9				
Effluent	5	106±6	0.3	1.1	33	84±7	0.3	0.8	37	30
	50	92±4				89±3				
	100	109±7				85±13				

TABLE S2 Concentrations (ngL⁻¹) of BZT and MeBZT in influent and effluent wastewater determined in each studied WWTP.

TABLE S2 Concentrations (ngL⁻¹) of BZT and MeBZT in influent and effluent wastewater determined in each studied WWTP.

WWTPs	Concentrations (ngL ⁻¹)			
	BZT		MeBZT	
	Influent	Effluent	Influent	Effluent
LLE	177.6	114.1	1289.0	1136.5
REU	3300.0	419.3	5406.6	2023.5
TAR	215.7	240.2	1341.8	1400.0
VDP	2097.0	504.6	3626.4	1329.4
VSS	76.5	26.7	778.7	676.5
LLA	855.0	214.1	8307.7	3576.5
SRS	2466.0	1513.8	6857.1	4682.4
GIR	768.0	258.7	2195.6	1400.0
MAT	474.0	122.9	2587.9	1729.4
MDV	1335.0	311.9	1681.3	1882.4
VTC	126.9	204.3	3560.4	2247.1
GAV	861.0	45.8	2096.7	587.1
MON	1092.0	383.5	3059.3	1482.4
PRA	1356.0	364.2	12857.1	3035.3
RUB	4380.0	583.5	47142.9	10541.2
SFL	672.0	324.8	3659.3	1435.3
BES	1029.0	362.4	3428.6	2117.6
GRA	1092.0	423.9	2254.9	1976.5
MAN	1476.0	250.0	3293.4	687.1
TEI	519.0	182.6	2465.9	1858.8

TABLE S3 Occurrence and removal rates of both BZTs in wastewater of this study and the literature.

		BZT				MeBZT						
		Concentrations ngL-1				Concentrations ngL-1						
		Influent		Effluent		Influent		Effluent				
Max	Min	Max	Min	Max	Min	Max	Min	Max	Min	Location	Reference	
4380	76.5	1513.8	26.7	-61 - 95	778.7	42900	587.1	10541.2	587.1	-12 - 79	Catalonia	This study
44000	17000	18000	7000	29 - 58	1100	4900	1000	1700	1000	19 - 69	Berlin	Reentma et al. 2010
73000	15000	100000	11000	-28 - 76	1100	5600	100	200	100	18 - 70	Switzerland	Voutsas et al. 2006
587	580	385	325	25 - 37	5156	5737	7735	15841	7735	26 - 68	Athens	Asimakopoulous et al. 2013

Información adicional - publicación Nº 5

Ecological risk assessment associated to the removal of endocrine-disrupting parabens and benzophenone-4 in wastewater treatment

Por

D. Molins-Delgado, M.S. Díaz-Cruz, D. Barceló

en

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SUPPORTING INFORMATION
for
ECOLOGICAL RISK ASSESSMENT ASSOCIATED TO THE
REMOVAL OF ENDOCRINE-DISRUPTING PARABENS
AND BENZOPHENONE 4 IN WASTEWATER TREATMENT

Daniel Molins-Delgado¹, M. Silvia Díaz-Cruz^{1*}, Damià Barceló^{1,2}

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Table A.1 Instrumental performance of the HPLC-MS/MS analytical method developed. Instrumental limit of detection (ILOD); instrumental limit of quantification (ILOQ); relative standard deviation (RSD).

Table A.2 Recovery rates (REC), method limits of detection (LOD) and quantification (LOQ) and matrix effects (ME%) in influent (a) and effluent (b) wastewater samples for the parabens and benzophenone 4.

Table A.3 Concentrations (ngL⁻¹) of parabens and BP4 in influent and effluent wastewater streams determined in each studied WWTP. <LOQ: below limit of quantification; n.d.: not detected.

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Table A.1: Instrumental performance of the HPLC-MS/MS analytical method developed. Instrumental limit of detection (ILOD); instrumental limit of quantification (ILOQ); relative standard deviation (RSD).

	Retention time (min)	Linearity (ngL ⁻¹)	Correlation coefficient (r ²)	ILOD (pg)	ILOQ (pg)	Precision (RSD%) (n=5)	
						Intraday	Interday
Methylparaben	7.45	3 – 100	0.998	0.01	0.02	5	15
Propylparaben	8.21	0.1 – 500	0.997	0.03	0.10	3	7
Butylparaben	8.52	0.1 – 500	0.999	0.08	0.26	4	13
Benzylparaben	8.50	0.1 – 500	0.999	0.01	0.03	3	15
BP4	6.25	5 – 500	0.991	0.44	1.47	7	16

Table A.2: Recovery rates (REC), method limits of detection (LOD) and quantification (LOQ) and matrix effects (ME%) in influent (a) and effluent (b) wastewater samples for the parabens and benzophenone 4.

a)

	REC \pm SD			LOD (ngL ⁻¹)	LOQ (ngL ⁻¹)	ME%
	30 ngL ⁻¹	50 ngL ⁻¹	100 ngL ⁻¹			
Methylparaben	94 \pm 2	93 \pm 7	95 \pm 14	2.1	7.1	11
Propylparaben	106 \pm 15	111 \pm 10	112 \pm 13	2.1	7.1	8
Butylparaben	102 \pm 11	101 \pm 5	107 \pm 7	2.1	7.1	6
Benzylparaben	105 \pm 3	108 \pm 7	103 \pm 6	2.1	7.1	6
BP4	101 \pm 5	99 \pm 5	102 \pm 5	4.8	15.9	22

b)

	REC \pm SD			LOD (ngL ⁻¹)	LOQ (ngL ⁻¹)	ME%
	30 ngL ⁻¹	50 ngL ⁻¹	100 ngL ⁻¹			
Methylparaben	92 \pm 6	95 \pm 15	96 \pm 11	0.58	1.9	1
Propylparaben	112 \pm 10	112 \pm 14	107 \pm 12	1.1	3.6	1
Butylparaben	108 \pm 6	109 \pm 5	109 \pm 5	0.11	0.37	1
Benzylparaben	112 \pm 5	104 \pm 10	103 \pm 2	0.05	0.27	1
BP-4	106 \pm 5	103 \pm 5	108 \pm 5	2.3	7.7	10

Table A.3 Concentrations (ngL^{-1}) of parabens and BP4 in influent and effluent wastewater streams determined in each studied WWTP. <LOQ: below limit of quantification; n.d.: not detected.

WWTPs	Benzy/paraben (ngL^{-1})		Butylparaben (ngL^{-1})		Propylparaben (ngL^{-1})		Methylparaben (ngL^{-1})		BP-4 (ngL^{-1})	
	Influent	Effluent	Influent	Effluent	Influent	Effluent	Influent	Effluent	Influent	Effluent
LLE	< LOQ	< LOQ	96	< LOQ	5010	< LOQ	2466	< LOQ	690	1080
REU	< LOQ	< LOQ	7.05	< LOQ	1371	< LOQ	< LOQ	< LOQ	1470	663
TAR	< LOQ	< LOQ	< LOQ	< LOQ	459	< LOQ	< LOQ	< LOQ	372	296
VDP	< LOQ	< LOQ	< LOQ	< LOQ	76.8	< LOQ	< LOQ	< LOQ	1062	277
VSS	< LOQ	< LOQ	39.3	< LOQ	858	< LOQ	83.6	137	1008	412
LLA	< LOQ	< LOQ	< LOQ	< LOQ	414	< LOQ	75.6	< LOQ	196.2	634
SRS	< LOQ	< LOQ	65.7	< LOQ	1695	< LOQ	2.58	< LOQ	1806	1060
GIR	< LOQ	< LOQ	16.35	< LOQ	1005	< LOQ	41.1	< LOQ	1332	585
MAT	< LOQ	< LOQ	< LOQ	< LOQ	696	< LOQ	801	< LOQ	1320	417
MDV	< LOQ	< LOQ	< LOQ	< LOQ	47.4	< LOQ	< LOQ	< LOQ	387	196
VIC	< LOQ	< LOQ	< LOQ	< LOQ	225.3	< LOQ	< LOQ	< LOQ	438	400
GAV	< LOQ	< LOQ	1.6	< LOQ	855	< LOQ	< LOQ	< LOQ	1317	487
MON	< LOQ	< LOQ	105	< LOQ	1494	< LOQ	< LOQ	< LOQ	1047	648
PRA	< LOQ	< LOQ	360	< LOQ	5700	< LOQ	2211	< LOQ	1074	720
RUB	< LOQ	< LOQ	< LOQ	< LOQ	156.6	< LOQ	< LOQ	< LOQ	516	321
SFL	< LOQ	< LOQ	< LOQ	< LOQ	363	< LOQ	< LOQ	< LOQ	969	376
BES	< LOQ	< LOQ	11.46	< LOQ	648	< LOQ	< LOQ	< LOQ	675	862
GRA	< LOQ	< LOQ	< LOQ	< LOQ	204.9	< LOQ	< LOQ	< LOQ	603	736
MAN	n.d.	< LOQ	71.1	< LOQ	2475	< LOQ	2220	< LOQ	984	86.1

Información adicional - publicación Nº 8

*Occurrence of organic UV filters and metabolites in lebranche mullet
(Mugil liza) from Brazil*

Por

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R.L. Ziolli, R.A. Hauser-Davis, E. Eljarrat, D. Barceló, M.S. Díaz-Cruz

en

Science of the Total Environment (enviado)

Supporting Information for

**OCCURRENCE OF ORGANIC UV FILTERS AND
METABOLITES IN LEBRANCHE MULLET (*Mugil liza*)
FROM BRAZIL**

Daniel Molins-Delgado, Ramón Muñoz, Sylvia Nogueira, Mariana B. Alonso, João Paulo Torres, Olaf Malm, Roberta Lourenço Ziolli, Rachel Ann Hauser-Davis, Ethel Eljarrat, Damià Barceló, M. Silvia Díaz-Cruz

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Table S7. Concentrations of each compound and the total load of UV-Fs ($\Sigma UV-Fs$) in each individual expressed in lipid weight (lw) in a) liver, b) muscle, and c) gill. <LOQ: below the limits of quantification; n.d.: not detected.

Table S1. Chemical name, abbreviation, CAS number, Log K_{ow}, and chemical structure for the target UV-Fs.

Name	Abbreviation	CAS number	Log K _{ow}	Structure
4-dihydroxybenzophenone	BP1	131-56-6	3.17	
2-hydroxy-4-methoxybenzophenone	BP3	131-57-7	3.79	
4-hydroxybenzophenone	4HB	1137-42-4	3.02	
4,4'-dihydroxybenzophenone	4DHB	611-99-4	2.55	
4-methylbenzylidene camphor	4MBC	36861-47-9	4.95	
Ethylhexyl dimethyl p-aminobenzoic	ODPABA	21245-02-3	6.15	
Ethylhexyl methoxycinnamate	EHMC	5466-77-3	5.80	
Octocrylene	OC	6197-30-4	7.53	

Table S2. Fresh and sample tissue weight, length and sampling site for each *Mugil liza* individual.

Individual CODE	Individual data		Sampling site	Sample weight		
	Weight (g)	Length (cm)		Gills (g dw)	Muscle (g dw)	Liver (g dw)
380	593.55	42.5	Ipiranga	0.52	0.53	0.5
389	721.79	45.0		0.51	0.55	0.56
392	662.33	43.0		0.53	0.57	0.57
399	932.45	47.0		0.51	0.54	0.54
404	678.00	47.0	Itaipu	0.51	0.53	0.53
406	811.47	46.5		0.45	0.54	0.53
407	900.00	46.9		0.46	0.51	0.57
408	543.05	39.5		0.51	0.55	0.5
409	803.31	47.6		0.52	0.55	0.58
413	935.47	46.5		0.54	0.53	0.59
415	625.84	43.0		0.53	0.53	0.54

Table S3. SRM transitions of the target compounds. E_{con} : cone energy; E_{col} : collision energy.

Compound	SRM Transition	E_{con} (V)	E_{col} (V)
BP1	215 → 137	40	27
	→ 105		29
BP3	229 → 151	40	25
	→ 105		27
4HB	199 → 121	40	25
	→ 105		27
4DHB	215 → 121	45	27
	→ 93		45
4MBC	255 → 212	61	29
	→ 105		41
OC	362 → 250	71	15
	→ 232		27
EHMC	291 → 179	51	25
	→ 161		13
ODPABA	279 → 166	86	27
	→ 151		43

Table S4. Linearity, instrumental limits of detection (LOD) and quantification (ILOQ), precision, recovery rates and methods limits of detection (LOD) and quantification (LOQ) for each compound.

Compound	Linearity	r ²	LOD (pg)	ILOQ (pg)	Precision (RSD%; n=7)		Recovery (%)	LOD (ngg ⁻¹ dw)	LOQ (ngg ⁻¹ dw)
					Intraday	Interday			
BP1	0.5-500	0.999	10	33	3	6	92	0.93	3.10
BP3	0.5-500	0.999	4	13	3	5	107	0.93	3.20
4HB	0.5-500	0.999	14	47	3	5	110	0.87	2.90
4DHB	0.5-500	0.999	14	47	3	5	96	0.93	3.10
4MBC	0.5-500	0.999	6	20	3	6	95	0.39	1.30
OC	0.5-500	0.999	10	33	5	8	75	0.39	1.30
EHMC	2.5-500	0.999	10	33	5	7	66	0.33	1.10
ODPABA	0.1-500	0.999	0.2	0.7	4	7	42	1.77	5.90

Table S5. Concentration of each compound in dry weight (dw) and the total amount of UV-Fs (Σ UV-Fs) in each individual in tissue of a) liver, b) muscle, and c) gills. <LOQ: below the limits of quantification; n.d.: not detected.

a)

Individual CODE	Concentrations of UV-Fs in liver (ngg ⁻¹ dw)								Σ UV-Fs
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	
380	9.51	26.1	139	101	<LOQ	13.7	<LOQ	<LOQ	289
389	4.41	13.7	49.9	92.2	9.53	6.84	<LOQ	5.03	182
392	17.1	74.4	131	14.8	9.12	7.89	<LOQ	25.9	280
399	<LOQ	11.8	83.7	61.7	7.14	7.16	<LOQ	<LOQ	172
404	14.1	50.6	77.6	154	<LOQ	<LOQ	<LOQ	<LOQ	296
406	10.1	8.30	78.0	57.5	6.92	4.63	n.d	<LOQ	165
407	4.95	10.8	118	67.1	98.8	6.84	n.d	7.53	314
408	<LOQ	20.9	113	5.77	<LOQ	11.7	<LOQ	5.03	156
409	<LOQ	7.55	5.47	64.2	<LOQ	7.16	<LOQ	<LOQ	84.4
413	<LOQ	8.77	36.6	34.9	<LOQ	6.74	<LOQ	<LOQ	87.0
415	3.71	32.6	66.9	451	14.0	7.26	<LOQ	11.6	587

b)

Individual CODE	Concentrations of UV-Fs in muscle (ngg ⁻¹ dw)								Σ UV-Fs
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	
380	<LOQ	12.1	3.25	6.40	49.4	9.26	<LOQ	57.8	138
389	<LOQ	3.5	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	5.24	8.74
392	<LOQ	3.91	<LOQ	17.4	<LOQ	5.16	<LOQ	<LOQ	26.5
399	<LOQ	15.4	3.15	16.6	17.1	23.4	<LOQ	24.9	101
404	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	n.a
406	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	n.a
407	n.d.	<LOQ	<LOQ	<LOQ	<LOQ	6.00	<LOQ	13.3	20.2
408	<LOQ	<LOQ	5.70	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	5.70
409	<LOQ	4.84	3.02	4.29	<LOQ	9.89	<LOQ	<LOQ	22.0
413	<LOQ	4.47	22.6	<LOQ	<LOQ	16.4	<LOQ	22.3	65.8
415	<LOQ	<LOQ	3.07	<LOQ	<LOQ	n.d.	<LOQ	n.d.	3.07

c)

Individual CODE	Concentrations of UV-Fs in gills (ngg ⁻¹ dw)								
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	ΣUV-Fs
380	<LOQ	6.62	9.45	23.5	<LOQ	6.21	<LOQ	9.96	55.7
389	<LOQ	3.33	12.1	19.4	<LOQ	5.79	<LOQ	9.85	50.5
392	<LOQ	3.07	6.45	17.1	<LOQ	7.47	<LOQ	2.40	36.5
399	<LOQ	<LOQ	18.2	6.59	<LOQ	<LOQ	<LOQ	<LOQ	24.8
404	<LOQ	24.0	8.64	11.3	<LOQ	4.42	<LOQ	9.85	58.2
406	<LOQ	9.42	9.45	15.3	7.27	13.3	<LOQ	10.7	65.4
407	<LOQ	<LOQ	7.27	3.86	<LOQ	7.16	<LOQ	6.20	24.5
408	<LOQ	<LOQ	5.28	3.29	<LOQ	4.95	<LOQ	<LOQ	13.5
409	<LOQ	6.80	31.64	13.7	<LOQ	14.5	<LOQ	16.4	83.0
413	<LOQ	3.74	16.36	8.76	<LOQ	4.63	<LOQ	16.8	50.3
415	<LOQ	3.26	10.45	11.4	<LOQ	6.21	<LOQ	9.59	41.0

Table S6. Concentrations of each compound in wet weight (ww) and the total load of UV-Fs (Σ UV-Fs) in each individual in a) liver, b) muscle, and c) gill. <LOQ: below the limits of quantification; n.d.: not detected.

a)

Individual CODE	Concentrations of UV-Fs in liver (ngg ⁻¹ ww)								
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	Σ UV-Fs
380	2.9	7.8	41.7	30.3	<LOQ	4.1	<LOQ	<LOQ	86.8
389	1.3	4.1	15.0	27.7	2.9	2.1	<LOQ	1.5	54.5
392	5.1	22.3	39.4	4.4	2.7	2.4	<LOQ	7.8	84.1
399	<LOQ	3.5	25.1	18.5	2.1	2.1	<LOQ	<LOQ	51.5
404	4.2	15.2	23.3	46.2	<LOQ	<LOQ	<LOQ	<LOQ	88.9
406	3.00	2.5	23.4	17.3	2.1	1.4	n.d.	<LOQ	49.6
407	1.5	3.2	35.4	20.1	29.6	2.1	n.d.	2.3	94.2
408	<LOQ	6.3	33.9	1.7	<LOQ	3.5	<LOQ	1.5	46.9
409	<LOQ	2.3	1.6	19.3	<LOQ	2.1	<LOQ	<LOQ	25.3
413	<LOQ	2.6	11.0	10.5	<LOQ	2.0	<LOQ	<LOQ	26.1
415	1.1	9.8	20.1	135.3	4.2	2.2	<LOQ	3.5	176.1

b)

Individual CODE	Concentrations of UV-Fs in muscle (ngg ⁻¹ ww)								
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	Σ UV-Fs
380	<LOQ	3.6	1.0	1.9	14.8	2.8	<LOQ	17.3	41.5
389	<LOQ	1.1	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	1.6	2.6
392	<LOQ	1.2	<LOQ	5.2	<LOQ	1.5	<LOQ	<LOQ	7.9
399	<LOQ	4.6	0.9	5.0	5.1	7.0	<LOQ	7.5	30.2
404	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	0.0
406	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	0.0
407	n.d.	<LOQ	<LOQ	<LOQ	<LOQ	1.8	<LOQ	4.0	6.0
408	<LOQ	<LOQ	1.7	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	1.7
409	<LOQ	1.5	0.9	1.3	<LOQ	3.0	<LOQ	<LOQ	6.6
413	<LOQ	1.3	6.8	<LOQ	<LOQ	4.9	<LOQ	6.7	19.7
415	<LOQ	<LOQ	0.9	<LOQ	<LOQ	n.d.	<LOQ	n.d.	0.9

c)

Individual CODE	Concentrations of UV-Fs in gills (ngg ⁻¹ ww)								
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	ΣUV-Fs
380	<LOQ	2.0	2.8	7.1	<LOQ	1.9	<LOQ	3.0	16.7
389	<LOQ	1.0	3.6	5.8	<LOQ	1.7	<LOQ	3.0	15.1
392	<LOQ	0.9	1.9	5.1	<LOQ	2.2	<LOQ	0.7	10.9
399	<LOQ	<LOQ	5.5	2.0	<LOQ	<LOQ	<LOQ	<LOQ	7.4
404	<LOQ	7.2	2.6	3.4	<LOQ	1.3	<LOQ	3.0	17.5
406	<LOQ	2.8	2.8	4.6	2.2	4.0	<LOQ	3.2	19.6
407	<LOQ	<LOQ	2.2	1.2	<LOQ	2.1	<LOQ	1.9	7.3
408	<LOQ	<LOQ	1.6	1.0	<LOQ	1.5	<LOQ	<LOQ	4.1
409	<LOQ	2.0	9.5	4.1	<LOQ	4.4	<LOQ	4.9	24.9
413	<LOQ	1.1	4.9	2.6	<LOQ	1.4	<LOQ	5.0	15.1
415	<LOQ	1.0	3.1	3.4	<LOQ	1.9	<LOQ	2.9	12.3

Table S7. Concentrations of each compound in lipid weight (lw) and the total load of UV-Fs (Σ UV-Fs) in each individual in a) liver, b) muscle, and c) gill. <LOQ: below the limits of quantification; n.d.: not detected.

a)

Individual CODE	Concentrations of UV-Fs in liver (ngg ⁻¹ lw)								
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	Σ UV-Fs
380	77	212	1130	821	<LOQ	111	<LOQ	<LOQ	2352
389	36	111	406	750	77	56	<LOQ	41	1477
392	139	605	1067	120	74	64	<LOQ	211	2280
399	<LOQ	96	680	502	58	58	<LOQ	<LOQ	1394
404	115	411	631	1252	<LOQ	<LOQ	<LOQ	<LOQ	2409
406	82	67	634	467	56	38	n.d.	<LOQ	1345
407	40	88	959	546	803	56	n.d.	61	2553
408	<LOQ	170	919	47	<LOQ	95	<LOQ	41	1272
409	<LOQ	61	44	522	<LOQ	58	<LOQ	<LOQ	686
413	<LOQ	71	298	284	<LOQ	55	<LOQ	<LOQ	707
415	30	265	544	3667	114	59	<LOQ	94	4773

b)

Individual CODE	Concentrations of UV-Fs in muscle (ngg ⁻¹ lw)								
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	Σ UV-Fs
380	<LOQ	399	107	211	1630	306	<LOQ	1908	4561.4
389	<LOQ	116	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	173	288.4
392	<LOQ	129	<LOQ	574	<LOQ	170	<LOQ	<LOQ	873.6
399	<LOQ	508	104	548	564	772	<LOQ	822	3318.5
404	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	n.a
406	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	n.a.
407	n.d.	<LOQ	<LOQ	<LOQ	<LOQ	198	<LOQ	439	665.0
408	< LOQ	< LOQ	188	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	188.1
409	< LOQ	160	100	142	<LOQ	326	<LOQ	<LOQ	727.4
413	< LOQ	148	746	<LOQ	<LOQ	541	<LOQ	736	2170.6
415	< LOQ	< LOQ	101	<LOQ	<LOQ	n.d.	<LOQ	n.d.	101.3

c)

Individual CODE	Concentrations of UV-Fs in gills (ngg ⁻¹ lw)								
	BP1	BP3	4HB	4DHB	EHMC	4MBC	ODPABA	OC	ΣUV-Fs
380	<LOQ	218	312	776	<LOQ	205	<LOQ	329	1840
389	<LOQ	110	399	640	<LOQ	191	<LOQ	325	1666
392	<LOQ	101	213	564	<LOQ	247	<LOQ	79	1204
399	<LOQ	<LOQ	601	217	<LOQ	<LOQ	<LOQ	<LOQ	818
404	<LOQ	792	285	373	<LOQ	146	<LOQ	325	1921
406	<LOQ	311	312	505	240	439	<LOQ	353	2160
407	<LOQ	<LOQ	240	127	<LOQ	236	<LOQ	205	808
408	<LOQ	<LOQ	174	109	<LOQ	163	<LOQ	<LOQ	446
409	<LOQ	224	1044	452	<LOQ	479	<LOQ	541	2741
413	<LOQ	123	540	289	<LOQ	153	<LOQ	554	1660
415	<LOQ	108	345	376	<LOQ	205	<LOQ	317	1350

Información adicional - publicación Nº 9

*A potential new threat to wild life:
presence of UV filters in bird eggs from a preserved area*

Por

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en

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Supporting Information for
**A POTENTIAL NEW THREAT TO WILD LIFE:
PRESENCE OF UV FILTERS IN BIRD EGGS FROM A
PRESERVED AREA**

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A.1. Sample pre-treatment and analysis

The method was optimised using hen eggs purchased from a local market. 0.1 g of the homogenized whole egg (without eggshell) were placed in beakers, completely covered with acetone and spiked with 50 ng of the surrogate standard solution of BP-¹³C. The acetone was left to evaporate at room temperature overnight, thus the eggs samples were dry prior analysis.

UV-Fs extraction was carried out by pressurised liquid extraction (PLE) using an ASE 350 Accelerated Solvent Extractor (Dionex Corporation, Sunnyvale, CA, USA). The experiments were conducted as follows: a cellulose filter followed by 1 g of Florisil (previously heated at 130 °C for 24 h) was placed at the bottom of the PLE cells. The freeze-dried eggs spiked with 50 ng of the surrogate standards were mixed in the PLE extraction cells with Florisil in order to perform first in-cell purification. Extraction was achieved in 2 static cycles of 5 min each at 100 °C and 1500 psi using the mixture AcEt/DCM (1:1, v/v) as extracting solvent.

The PLE extracts were diluted to 200 mL with water (to keep MeOH < 5%), and further purified by solid phase extraction (SPE) using Isolute C18 cartridges, from Biotage. The cartridges were conditioned with 5 mL of the mixture AcEt/DCM (1:1, v/v) followed by 5 mL of MeOH and 5 mL of water. Then the PLE diluted extracts were loaded onto the SPE cartridges using a Baker vacuum system (J.T. Baker, The Netherlands). Further, the cartridges were washed with 5 mL of water and dried. Then, the compounds retained in the SPE cartridges were eluted with 7 mL of the mixture AcEt/DCM (1:1, v/v) and 2 mL of DCM. Finally, the joint extracts were evaporated with nitrogen and reconstituted with 1 mL of the internal standards solution containing BP3-d₅, 4MBC-d₄, and AllylBZT at 20 ngmL⁻¹ in ACN. Analyses were carried out in triplicate. The lipid content was determined gravimetrically.

UV-Fs' separation and detection were performed by liquid chromatography-tandem mass spectrometry using a Symbiosis Pico chromatograph from Spark Holland (Emmen, The Netherlands) attached to a 4000 Q TRAPTM MS/MS analyser from Applied Biosystems-Sciex (Foster City, California, USA) (HPLC-QqLIT-MS/MS). The chromatographic separation was achieved on a Hibar Purospher[®] STAR[®] HR R-18 ec. (50 mm × 2.0 mm, 5 µm) from Merck, preceded by a guard column of the same packaging material. The mobile phase consisted of HPLC-grade water and ACN, both 0.1% formic acid. The total run time was 20 min. The mobile phase flow rate and the injection volume were set to 0.3 mL min⁻¹ and 20 µL, respectively. The chromatographic gradient started at 5% organic phase and reached 25% during the next 6 min., 4 min. later the gradient reached 100%, keeping it constant for 8 min. At minute 18 the gradient went back to the initial conditions.

MS/MS detection was performed in positive electrospray ionization (ESI+) under selected reaction monitoring (SRM) mode for enhanced selectivity and sensitivity. The two most intense transitions from the precursor molecular ion ((M+H)⁺) were monitored per compound. The most abundant transition was used for quantification, whereas the other one was used for confirmation. Instrument control, data acquisition and processing were performed with Analyst 1.4.2 software from Applied Biosystems/MDS Sciex.

Table S1. Name, abbreviation, CAS number and Log K_{ow} of the studied UV filters. In italic, metabolites of BP3.

Name	Abbreviation	CAS number	Log K_{ow}
<i>Benzophenones</i>			
2,4-Dihydroxybenzophenone	BP1	131-56-6	3.17
2-Hydroxy-4-methoxybenzophenone (oxybenzone)	BP3	131-57-7	3.79
4-Hydroxybenzophenone	4HB	1137-42-4	3.02
4,4'-Dihydroxybenzophenone	4DHB	611-99-4	2.55
<i>Crylenes</i>			
2-Ethylhexyl-2-cyano-3,3-diphenylacrylate	OC	6197-30-4	7.53
<i>p-aminobenzoic acid derivatives</i>			
2-Ethylhexyl-4-(dimethyl-amino) benzoate	ODPABA	21245-02-3	6.15
<i>Benzotriazole derivatives</i>			
2-(2-Benzotriazolyl)-p-cresol	UVP	2440-22-4	4.3

Table S2. Sample name, species, $\delta^{13}\text{C}$, $\delta^{15}\text{N}$, water content (%H₂O) and total lipids content (%lipid) per sample.

Sample Name	Order	Common name	Scientific name	$\delta^{13}\text{C}$	$\delta^{15}\text{N}$	%H ₂ O	%lipid*
27e		Western marsh harrier	Circus aeruginosus	-23.86	14.98	75.6	24.4
28e				-23.99	8.64	81.7	---
29e				-23.75	8.91	82.1	32.8
30e				-23.60	8.77	81.0	27.2
34e				-22.38	7.96	81.9	30.5
42e				-23.99	9.53	79.1	21.4
43e				-24.09	8.92	79.4	23.8
44e				-24.29	8.58	76.7	10.6
45e				-23.73	11.44	71.3	29.6
46e				-23.41	11.45	78.8	25.6
47e				-22.88	11.29	79.7	29.5
1e				-21.80	9.38	77.6	33.3
2e				-22.09	8.84	76.5	31.5
3e				-21.55	9.04	76.7	18.3
4e				-22.18	8.81	76.8	21.6
5e				-23.90	11.78	76.3	32.0
6e				-23.34	10.48	76.3	33.8
7e				-23.34	10.94	80.8	26.9
8e				-22.90	9.62	76.8	31.0
14e		Slender-billed gull	Chrococephalus genei	-18.87	12.37	73.4	36.9
19e				-19.05	9.47	72.2	36.8
21e				-17.51	11.42	72.7	36.2
23e				-17.62	10.22	70.8	42.5
24e				-18.02	11.39	74.2	33.8
52e				-22.96	14.20	67.1	37.7
54e				-23.68	13.17	72.3	18.6
57e				-22.11	12.34	73.6	47.1
59e				-23.92	15.07	69.3	41.9
61e				-18.42	15.85	29.5	42.6
64e				-20.46	13.04	31.0	45.2
65e				-20.20	14.51	32.1	41.0
66e				-16.05	14.54	31.9	39.2
67e				-20.15	14.59	33.4	35.3
68e				-16.72	15.70	30.3	46.9
69e				-19.20	14.04	32.2	44.2
70e				-20.07	13.00	32.5	40.5
71e				-20.92	13.32	32.8	46.3
72e				-16.55	15.68	32.7	45.2
73e				-19.14	14.81	34.2	46.1

*lipids determined in dry-frozen samples.

Table S3. HPLC-MS/MS instrumental performance, recovery rates and method limit of detection (MLOD) and quantification (MLOQ). ILOD: instrumental limits of detection; ILOQ: instrumental limit of quantification. RSD: relative standard deviation.

Compound	Calibration range (ng g ⁻¹ dw)	r ²	ILOD (pg)	ILOQ (pg)	Precision (RSD)		Recovery rate (%)		MLOD (ng g ⁻¹ dw)	MLOQ (ng g ⁻¹ dw)
					Intraday	Inteday	25 ng g ⁻¹ dw	50 ng g ⁻¹ dw		
BP3	0.1-500	0.999	0.66	2.21	3	11	74 ± 5	77 ± 2	0.29	0.97
BP1	0.1-500	0.999	5.66	18.97	5	7	46 ± 3	39 ± 2	0.25	0.85
4HB	0.1-500	0.999	3.18	10.60	5	10	45 ± 4	57 ± 3	0.53	1.77
4DHB	0.1-500	0.999	0.54	1.80	1	5	54 ± 3	55 ± 2	0.15	0.51
ODPABA	0.1-500	0.999	0.14	0.49	7	9	54 ± 11	39 ± 5	0.04	0.13
OC	0.1-500	0.999	0.63	2.12	9	15	107 ± 17	90 ± 7	0.26	0.86
UVP	0.1-500	0.999	0.14	0.49	10	18	125 ± 8	124 ± 10	0.13	0.43

Table S4. Concentrations of UV-Fs expressed in ngg⁻¹ dw of freeze-dried egg each sample. <MLOQ: below the limit of quantification, n.d.: not detected.

Sample Name	Order	Common name	Scientific name	Concentration (ng g ⁻¹ dw)									
				BP1	BP3	4HB	4DHB	ODPABA	OC	UVP			
27e		Western marsh harrier	Circus aeruginosus	40.6	46.7	895	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	2.7	
28e	40.4			27.2	267	<MLOQ	<MLOQ	<MLOQ	3.0				
29e	53.5			18.3	70.6	<MLOQ	<MLOQ	<MLOQ	0.4				
30e	28.0			24.9	20.0	<MLOQ	<MLOQ	<MLOQ	1.0				
34e	36.4			25.5	48.1	<MLOQ	<MLOQ	<MLOQ	<MLOQ				
42e	27.9			35.2	82.9	<MLOQ	<MLOQ	<MLOQ	<MLOQ				
43e	38.1			21.3	26.2	132	<MLOQ	<MLOQ	<MLOQ				
44e	42.7			23.7	1,200	<MLOQ	<MLOQ	<MLOQ	<MLOQ				
45e	42.3			27.0	n.d.	<MLOQ	<MLOQ	<MLOQ	<MLOQ				
46e	47.4			18.5	153	<MLOQ	<MLOQ	<MLOQ	<MLOQ				
47e	36.2			26.0	27.3	<MLOQ	<MLOQ	<MLOQ	<MLOQ				
1e	<MLOQ			20.6	104	<MLOQ	<MLOQ	<MLOQ	26.7	<MLOQ			
2e	<MLOQ			19.5	224	<MLOQ	<MLOQ	<MLOQ	15.2	<MLOQ			
3e	<MLOQ			29.2	1,059	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
4e	<MLOQ			25.6	360	<MLOQ	<MLOQ	<MLOQ	<MLOQ	n.d.			
5e	<MLOQ			19.8	408	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
6e	<MLOQ			20.9	3,348	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
7e	<MLOQ	22.0	210	<MLOQ	<MLOQ	n.d.	<MLOQ	<MLOQ					
8e	<MLOQ	20.7	378	29.04	<MLOQ	<MLOQ	<MLOQ	<MLOQ					
14e		Slender-billed gull	Chrococephalus genei	<MLOQ	22.5	266	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
19e	<MLOQ			24.7	86.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	n.d.			
21e	594			44.4	72.8	<MLOQ	<MLOQ	<MLOQ	23.8	<MLOQ			
23e	677			38.5	111	<MLOQ	<MLOQ	<MLOQ	65.2	8.4			
24e	27.8			31.4	83.8	<MLOQ	<MLOQ	<MLOQ	11.4	1.4			
52e	66.1			49.3	472	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
54e	34.9			18.2	35.8	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
57e	29.0			21.2	12.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	1.8			
59e	39.9			24.3	37.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	2.0			

Table S4. Continued.

Sample Name	Order	Common name	Scientific name	Concentration (ng g ⁻¹ dw)									
				BP1	BP3	4HB	4DHB	ODPABA	OC	UVP			
61e				41.0	17.1	560	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
64e				87.3	16.9	332	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
65e				24.9	17.3	66.2	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
66e				23.3	23.7	18.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
67e				37.1	22.6	13.5	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
68e				27.2	18.5	24.5	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
69e				30.7	22.4	41.9	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	0.2	
70e				33.4	31.2	107	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
71e				33.9	25.7	38.7	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
72e				50.3	21.9	120	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
73e				29.4	25.3	20.7	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	

Table S5. Concentrations of UV-Fs expressed in ngg⁻¹ lw of each freeze-dried egg sample. <MLOQ: below the limits of quantification. n.d.: not detected; n.a.: not applicable.

Sample Name	Order	Common name	Scientific name	Concentration (ng g ⁻¹ lw)									
				BP1	BP3	4HB	4DHB	ODPABA	OC	UVP			
27e		Western marsh harrier	<i>Circus aeruginosus</i>	167	191	3,670	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	11.2	
28e	n.a.			n.a.	n.a.	n.a.	n.a.	n.a.	n.a.				
29e	163			55.8	215	<MLOQ	<MLOQ	<MLOQ	<MLOQ	1.25			
30e	103			91.7	73.5	<MLOQ	<MLOQ	<MLOQ	<MLOQ	3.7			
34e	119			83.4	158	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
42e	130			165	387	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
43e	160			89.7	110	553	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
44e	403			223	11,318	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
45e	143			91.1	n.d.	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
46e	185			72.3	596	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
47e	123			88.3	92.5	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
1e	<MLOQ			61.7	311	<MLOQ	<MLOQ	<MLOQ	26.6	<MLOQ			
2e	<MLOQ			61.9	711	<MLOQ	<MLOQ	<MLOQ	15.2	<MLOQ			
3e	<MLOQ			160	5,785	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
4e	<MLOQ			119	1,668	<MLOQ	<MLOQ	<MLOQ	<MLOQ	n.d.			
5e	<MLOQ			62.0	1,274	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
6e	<MLOQ			61.7	9,905	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
7e	<MLOQ	81.8	780	<MLOQ	n.d.	<MLOQ	<MLOQ	<MLOQ					
8e	<MLOQ	66.8	1,221	93.7	<MLOQ	<MLOQ	<MLOQ	<MLOQ					
14e		Slender-billed gull	<i>Chrocephalus genei</i>	<MLOQ	61.0	722	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
19e	<MLOQ			67.1	231	<MLOQ	<MLOQ	<MLOQ	<MLOQ	n.d.			
21e	1,642			123	201	<MLOQ	<MLOQ	<MLOQ	65.8	<MLOQ			
23e	1,594			90.6	262	<MLOQ	<MLOQ	<MLOQ	153	19.7			
24e	82.2			93.0	248	<MLOQ	<MLOQ	<MLOQ	11.4	1.4			
52e	175			131	1,252	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
54e	187			98.1	193	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ			
57e	61.5			45.0	25.5	<MLOQ	<MLOQ	<MLOQ	<MLOQ	2.5			
59e	95.2			58.0	88.2	<MLOQ	<MLOQ	<MLOQ	<MLOQ	4.7			

Table S5. Continued.

Sample Name	Order	Common name	Scientific name	Concentration (ng g ⁻¹ fw)							
				BP1	BP3	4HB	4DHB	ODPABA	OC	UVP	
61e				96.4	40.8	1,315	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
64e				193	37.3	735	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
65e				60.8	42.1	161	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
66e				59.3	60.5	46.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
67e				105	64.1	38.2	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
68e				58.1	39.5	52.4	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
69e				69.4	50.6	94.7	<MLOQ	<MLOQ	<MLOQ	<MLOQ	0.4
70e				82.4	77.7	265	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
71e				73.3	55.5	83.7	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
72e				111	48.5	266	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
73e				63.8	54.8	44.9	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ

Table S6. Concentrations of UV-Fs expressed in ngg⁻¹ ww of each freeze-dried egg sample. <MLOQ: below the limit of quantification. n.d.: not detected.

Sample Name	Order	Common name	Scientific name	Concentration (ng g ⁻¹ ww)								
				BP1	BP3	4HB	4DHB	ODPABA	OC	UVP		
27e		Western marsh harrier	<i>Circus aeruginosus</i>	9.9	11.4	219	<MLOQ	<MLOQ	<MLOQ	n.d.	0.7	
28e	7.4			5.0	48.8	<MLOQ	<MLOQ	<MLOQ	<MLOQ	0.5		
29e	9.6			3.3	12.7	<MLOQ	<MLOQ	<MLOQ	<MLOQ	0.1		
30e	5.3			4.7	3.8	<MLOQ	<MLOQ	<MLOQ	<MLOQ	0.2		
34e	6.6			4.6	8.7	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
42e	5.8			7.4	17.3	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
43e	7.9			4.4	5.4	27.2	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
44e	10.0			5.5	280	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
45e	12.2			7.7	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
46e	10.1			3.9	32.4	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
47e	7.3			5.3	5.5	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
1e						<MLOQ	4.6	23.3	<MLOQ	<MLOQ	6.0	<MLOQ
2e						<MLOQ	4.6	52.6	<MLOQ	<MLOQ	3.6	<MLOQ
3e						<MLOQ	6.8	247	<MLOQ	<MLOQ	<MLOQ	<MLOQ
4e				<MLOQ	5.9	83.5	<MLOQ	<MLOQ	<MLOQ	n.d.		
5e				<MLOQ	4.7	96.8	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
6e				<MLOQ	5.0	795	<MLOQ	<MLOQ	<MLOQ	<MLOQ		
7e				<MLOQ	4.2	40.4	<MLOQ	n.d.	<MLOQ	<MLOQ		
8e				<MLOQ	4.8	87.9	6.7	<MLOQ	<MLOQ	<MLOQ		
14e		Slender-billed gull	<i>Chroicocephalus genei</i>	<MLOQ	6.0	71.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
19e				<MLOQ	6.9	23.9	<MLOQ	<MLOQ	<MLOQ	<MLOQ	n.d.	
21e				162	12.1	19.9	<MLOQ	<MLOQ	<MLOQ	6.5	<MLOQ	
23e				198	11.2	32.4	<MLOQ	<MLOQ	<MLOQ	19.0	2.4	
24e				7.2	8.1	21.7	<MLOQ	<MLOQ	<MLOQ	3.0	0.4	
52e				21.8	16.2	155	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
54e				9.7	5.1	9.9	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	
57e				7.6	5.6	3.2	<MLOQ	<MLOQ	<MLOQ	<MLOQ	0.3	
59e				12.2	7.4	11.3	<MLOQ	<MLOQ	<MLOQ	<MLOQ	0.6	

Table S6. Continued.

Sample Name	Order	Common name	Scientific name	Concentration (ng g ⁻¹ ww)								
				BP1	BP3	4HB	4DHB	ODPABA	OC	UVP		
61e				28.9	12.3	395	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
64e				60.2	11.6	229	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
65e				16.9	11.7	44.9	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
66e				15.8	16.2	12.2	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
67e				24.7	15.1	9.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
68e				19.0	12.9	17.1	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
69e				20.8	15.2	28.4	<MLOQ	<MLOQ	<MLOQ	<MLOQ	0.1	<MLOQ
70e				22.6	21.3	72.4	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
71e				22.8	17.3	26.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
72e				33.8	14.8	81.0	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ
73e				19.4	16.6	13.6	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ	<MLOQ

Table S7. Total mean concentrations (expressed in ng g⁻¹ lw) of PBDEs, dechloranes, pyrethroids, and UV-Fs in the studied species from Doñana. n.a.: not applicable. SD, standard deviation.

Order	Common Name	Species	∑PBDE ¹	SD	∑Dechloranes ¹	SD	∑Pyrethroid ²	SD	∑UV-Fs	SD
	Western marsh harrier	Circus aeruginosus	23.4	n.a.	161	n.a.	4.88	n.a.	4,046.80	n.a.
	Common kestrel	Falco tinnunculus	12.7	9.37	8.88	7.18	6.17	9.7	397.91	3,639.06
Ciconiiformes	White stork	Ciconia ciconia	80	16	66.1	57.6	31.4	39	1,357.80	3,371.27
	Slender-billed gull	Chrococephalus genei	5.03	1	39.4	11.6	3.35	2	782.80	n.a.
	Black-headed gull	Chrococephalus ridibundus	5.98	1.99	63.4	30.5	162	128	1,247.35	981.46
	Gull-billed tern	Gelochelidon nilotica	6.62	3.19	23.6	16.6	61.5	80	361.72	651.16
Anseriformes	Gadwall	Anas strepera	5.66	1.2	5.93	1.33	5.79	3.4	215.20	415.15

¹ Barón, E.; Máfiez, M.; Andreu, A. C.; Sergio, F.; Hiraldo, F.; Eijarar, E.; Barceló, D. Bioaccumulation and biomagnification of emerging and classical flame retardants in birds of 14 species from Doñana Natural Space and surrounding areas (South-western Spain). Environ. Int. 2014, 68, 118-126.

² Corcellas, C.; Andreu, A.; Máfiez, M.; Sergio, F.; Hiraldo, F.; Eijarar, E.; Barceló, D. Pyrethroid insecticides in wild bird eggs from a World Heritage Listed Park: A case study in Doñana National Park (Spain). Environ. Pollut. 2017, 228, 321-330.

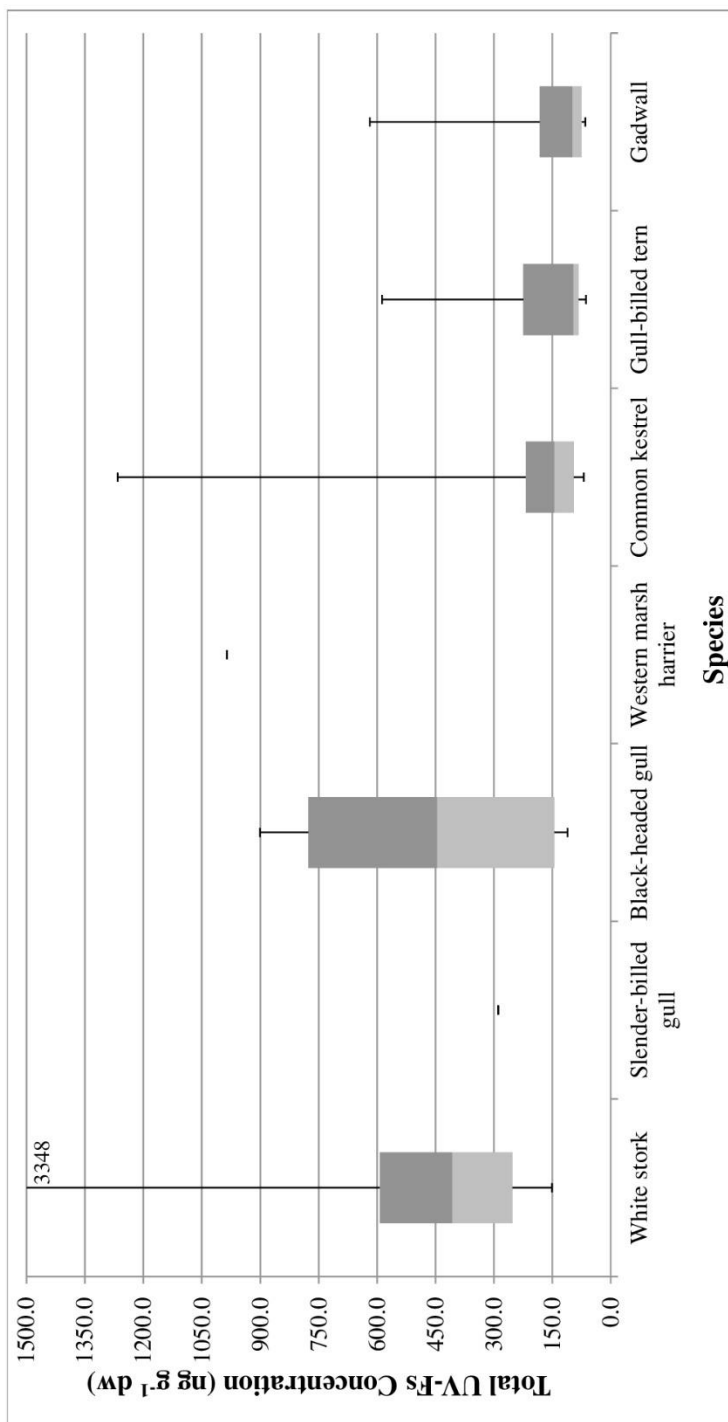


Figure S1. Total concentrations of UV-Fs in the studied eggs from wild bird species.

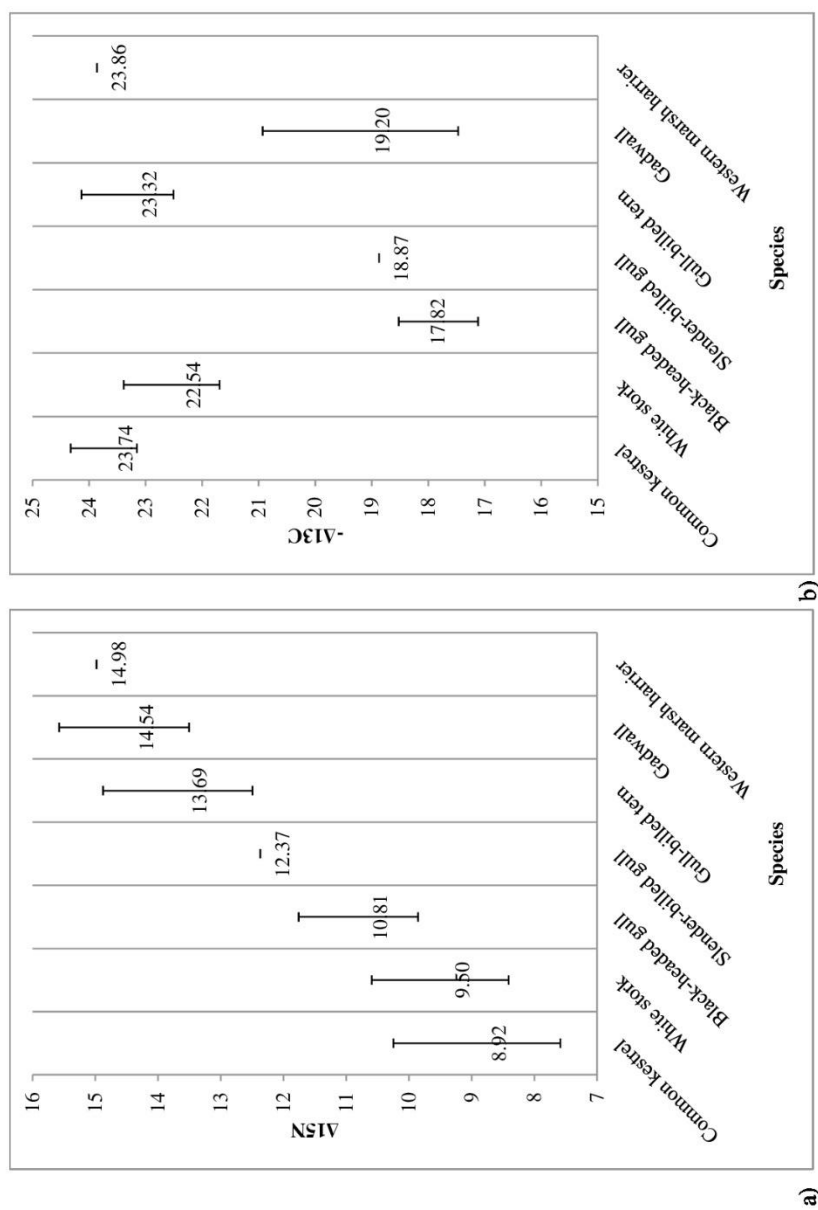


Figure S2. $\delta^{15}\text{N}$ (a) and $\delta^{13}\text{C}$ (b) ranges for the eggs from wild bird species. Mean value is indicated in each range.

Información adicional - publicación Nº 10

Determination of UV filters in human breast milk using turbulent flow chromatography and babies' daily intake estimation

Por

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en

Environmental Research (enviado)

Supporting information
for
**Determination of UV filters in human breast milk
using turbulent flow chromatography and
babies' daily intake estimation**

Daniel Molins-Delgado, María del Mar Olmo-Campos, Gemma Valeta-Juan, Vanessa Pleguezuelos-Hernández, Damià Barceló, M. Silvia Díaz-Cruz.

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Table A8. Concentrations of UV-Fs (ng g^{-1} lw milk) in the sample where UV-Fs were detected. <LOQ: below the limits of quantification; n.d.; not detected. In parenthesis, the value of the corresponding LOQ expressed in (ng g^{-1} lw milk).

Figure A1.

1a. Focus-Mode: Loading step. The sample is injected onto the TFC column, where the analyte is retained whereas the debris from the matrix is led to waste.

1b. Focusing: Transfer step. The flow of both columns is combined and led into the analytical column.

1c. Focus-Mode: Eluting Step. The analytes eluted from the TFC column into the analytical column. The TFC column is washed and conditioned for the next injection.

Table A1. Target compounds name, abbreviation, CAS number, and Log K_{ow}.

Compound name	Abbreviation	CAS N.	Log K _{ow}
Benzophenone derivatives			
2-Hydroxy-4-methoxybenzophenone	BP3	131-57-7	3.79
2,4-Dihydroxybenzophenone	BP1	131-56-6	3.14
4-Hydroxybenzophenone	4HB	1137-42-4	3.02
4,4'-Dihydroxybenzophenone	4DHB	611-99-4	2.55
Camphor derivatives			
4-Methylbenzylidene camphor	4MBC	36861-47-9	4.95
p-aminobenzoic acid derivatives			
Ethylhexyl dimethyl-p-aminobenzoic acid	ODPABA	21245-02-3	6.15
Ethyl p-aminobenzoic acid	EPABA	94-09-7	1.86
Cinamates			
Ethylhexyl methoxycinamate	EHMC	5466-77-3	5.8
Crylenes			
Octocrylene	OC	6197-30-4	7.53
Benzotriazole derivatives			
2-(5-tert-Butyl-2-hydroxyphenyl)benzotriazole	TBHPBT	3147-76-0	3.24
2-(2-Hydroxy-5-methylphenyl)benzotriazole	UVP	2440-22-4	4.3
2-(2-H-benzotriazol-2-yl)-4,6-bis(1-methyl-1-phenylethyl)phenol	UV234	70321-86-7	7.67
2-(2'-Hydroxy-3',5'-di-tert-butylphenyl)benzotriazole	UV320	3846-71-7	5.3
2-tert-butyl-6-(5-chloro-2-H-benzotriazol-2-yl)-4-methylphenol	UV326	3896-11-5	7.2
2,4-di-tert-butyl-6-(5-chloro-2-H-benzotriazol-2-yl)phenol	UV327	3864-99-1	6.6
2-(2-H-benzotriazol-2-yl)-4,6-di-tert-pentylphenol	UV328	25973-55-1	7.8
2-(2-H-Benzotriazol-2-yl)-4-(1,1,3,3-tetramethylbutyl)phenol	UV329	3147-75-9	6.21
Benzotriazole	BZT	95-14-7	1.23
Methyl benzotriazole	MeBZT	136-85-6	1.89

Table A2. Summary of the information present in the questionnaires handed out to the mothers.

Participating mothers	79
Sampling campaign	April-October 2015
Mother 's age	34.4±4.0
Mothers with allergies	29%
Mothers who drink coffee	33%
Coffee consumption (per day)	1.2±0.4
Mothers who drink infusions	3%
Vegetarian diet (partially)	1%
Pregnancies	
Primiparous	90%
Multiparous	10%
Birth procedure	
Natural	77%
Cesarean	23%
Baby sex	
Male	51%
Female	49%
Weight at birth (g)	3170±734
Gestation (weeks)	38±4
Lactation (months)	7±6

Table A3. Summary of the macronutrient analysis of the milk samples and related parameters. SNF: non-lipid solids.

Fat content (%)	3.7 ± 1.0
SNF (%)	8.1 ± 0.3
Lactose (%)	6.6 ± 0.3
Proteins (%)	1.2 ± 0.1
Density (kg m⁻³)	27.5 ± 1.4
pH	6.6 ± 0.2

Table A4. Turbulent Flow and Liquid Chromatographic experimental conditions. f.a.: formic acid.

Time (min)	Turbulent Flow Chromatography			Liquid Chromatography			
	Flow (ml min ⁻¹)	%Water (0.1% f.a.)	%MeOH (0.1% f.a.)	Status	Flow (ml min ⁻¹)	%Water (0.1% f.a.)	%MeOH (0.1% f.a.)
00:40	1	75	25	Loading Sample	0.3	75	25
02:40	0.2	0	100	Transfer to the chromatograph	0.2	50	50
04:10	1	50	50	Washing	0.3	10	90
04:25	1	0	100	Washing	0.3	0	100
14:00	1	0	100	Washing / Loading Loop	0.3	0	100
16:00	1	75	25	Conditioning	0.3	75	25
18:00	1	75	25	Conditioning	0.3	75	25

Table A5. Transitions, ion guide stacked ring energy (S-Lens), and collision energy (CE) for each compound.

Compound	Precursor ion m/z	Product ion m/z	S-Lens (V)	CE (V)
	215	137	61	18
	215	81	61	33
	229	151	76	24
	229	77	76	35
	199	121	54	17
	199	77	54	31
	215	121	62	18
	215	93	62	29
	255	165	67	38
	255	141	67	37
	291	161	30	63
	291	133	30	63
	362	232	71	21
	362	204	34	34
	278	151	119	30
	278	166	119	20
	166	120	60	18
	166	94	60	17
	120	92	76	21
	120	65	76	15
	134	79	78	24
	134	77	78	17
	268	212	73	19
	268	57	73	23
	226	120	54	30
	226	107	54	15
	448	370	117	22
	448	91	117	19
	324	212	113	18
	324	57	113	28
	316	260	74	18
	316	107	74	24
	358	302	69	17
	358	246	69	34
	352	282	88	13
	352	43	88	23
	324	167	60	47
	324	92	60	41

Table A6. Recovery rates at three spike levels for the different columns tested: a) Cyclone; b) Cyclone P; c) Cyclone MAX; d) C18 XL. n.a. not applicable-not results.

a)

Compound	Recovery rates (%) (n=7)					
	Spike level 100 ngg ⁻¹	RSD (%)	Spike level 50 ngg ⁻¹	RSD (%)	Spike level 25 ngg ⁻¹	RSD (%)
Cyclone						
BP1	82	5	71	1	67	1
BP3	79	4	69	1	54	6
4HB	79	2	71	2	62	2
4DHB	67	3	70	6	57	2
4MBC	66	3	61	8	61	3
ODPABA	77	6	64	6	63	4
EtPABA	54	7	53	8	47	7
OC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
EHMC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
BZT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
MeBZT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
TBPHBT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UVP	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV234	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV320	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV326	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV327	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV328	18	4	39	13	57	4
UV329	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.

b)

Compound	Recovery rates (%) (n=7)					
	Spike level 100 ngg ⁻¹	RSD (%)	Spike level 50 ngg ⁻¹	RSD (%)	Spike level 25 ngg ⁻¹	RSD (%)
Cyclone P						
BP1	77	1	64	7	63	6
BP3	73	7	69	6	65	13
4HB	73	1	64	5	65	8
4DHB	69	3	61	4	69	7
4MBC	70	8	69	6	65	3
ODPABA	47	2	41	6	39	10
EtPABA	61	3	65	9	80	15
OC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
EHMC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
BZT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
MeBZT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
TBPHBT	51	6	34	4	35	15
UVP	62	2	58	9	27	14
UV234	45	2	51	4	67	5
UV320	37	5	39	8	45	11
UV326	11	5	13	9	13	3
UV327	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV328	2	18	3	12	3	18
UV329	44	2	42	7	35	1

c)

Recovery rates (%) (n =7)						
Compound	Spike level 100 ngg ⁻¹	RSD (%)	Spike level 50 ngg ⁻¹	RSD (%)	Spike level 25 ngg ⁻¹	RSD (%)
Cyclone MAX						
BP1	91	5	77	1	74	1
BP3	76	4	75	1	73	6
4HB	87	2	73	2	66	2
4DHB	177	3	66	6	10	2
4MBC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
ODPABA	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
EIPABA	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
OC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
EHMC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
BZT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
MeBZT	67	13	63	10	80	12
TBPHBT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UVP	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV234	37	5	49	14	59	15
UV320	37	17	54	10	60	6
UV326	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV327	53	6	86	8	180	3
UV328	33	6	68	6	125	5
UV329	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.

d)

Recovery rates (%) (n =7)						
Compound	Spike level 100 ngg ⁻¹	RSD (%)	Spike level 50 ngg ⁻¹	RSD (%)	Spike level 25 ngg ⁻¹	RSD (%)
C18 XL						
BP1	62	1	41	7	22	6
BP3	65	7	48	6	35	14
4HB	66	1	39	5	6	3
4DHB	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
4MBC	85	8	58	6	46	11
ODPABA	84	2	60	6	40	9
EtPABA	80	3	23	9	31	12
OC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
EHMC	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
BZT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
MeBZT	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
TBPHBT	75	5	75	7	83	12
UVP	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV234	46	6	53	6	74	1
UV320	34	1	27	8	37	7
UV326	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV327	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV328	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
UV329	55	7	54	6	56	14

Table A7. Concentrations of UV-Fs (ng g⁻¹ milk) in the samples where UV-Fs were detected. <LOQ: below the limits of quantification; n.d.; not detected. In parenthesis, the value of the corresponding LOQ expressed in (ng g⁻¹ milk).

Sample	UV-Fs concentrations (ng g ⁻¹ milk)							∑UV-Fs*
	BP3	4HB	4DHB	4MBC	TBHPBT	UV320	UV329	
14.10-07	n.d.	n.d.	<LOQ (2.0)	n.d.	n.d.	n.d.	n.d.	0.2
21.10-02	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	<LOQ (3.3)	2.1
21.10-05	<LOQ (0.30)	n.d.	n.d.	2.6	35.2	n.d.	n.d.	38.0
22.10-02	n.d.	n.d.	n.d.	n.d.	84.7	n.d.	n.d.	84.7
22.10-04	2.8	n.d.	n.d.	<LOQ (1.6)	n.d.	n.d.	n.d.	3.9
22.10-06	n.d.	n.d.	n.d.	<LOQ (1.6)	<LOQ (5.1)	n.d.	n.d.	4.4
23.10-06	<LOQ (0.3)	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	0.2
23.10-07	<LOQ (0.3)	n.d.	n.d.	n.d.	<LOQ (5.1)	n.d.	n.d.	3.5
23.10-10	<LOQ (0.3)	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	0.2
23.10-13	<LOQ (0.3)	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	0.2
28.10-03	<LOQ (0.3)	n.d.	n.d.	n.d.	n.d.	n.d.	98.9	99.1
28.10-04	<LOQ (0.3)	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	0.2
28.10-06	n.d.	n.d.	n.d.	n.d.	<LOQ (5.1)	n.d.	n.d.	3.3
28.10-08	n.d.	n.d.	n.d.	n.d.	<LOQ (5.1)	n.d.	n.d.	3.3
29.10-01	n.d.	n.d.	n.d.	n.d.	<LOQ (5.1)	n.d.	n.d.	3.3
29.10-06	n.d.	n.d.	n.d.	n.d.	<LOQ (5.1)	n.d.	n.d.	3.3
29.10-09	39.8	n.d.	19.9	n.d.	n.d.	n.d.	n.d.	59.7
29.10-10	52.1	3.9	23.3	n.d.	n.d.	n.d.	<LOQ (3.3)	81.4
29.10-11	40.2	n.d.	27.9	n.d.	n.d.	n.d.	116.9	185.0
29.10-12	226.5	n.d.	30.3	n.d.	n.d.	n.d.	n.d.	256.8
29.10-13	147.7	n.d.	43.3	n.d.	n.d.	346.0	434.1	971.1
30.10-01	76.1	n.d.	n.d.	n.d.	n.d.	523.6	186.5	786.1
30.10-02	68.7	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	68.6
30.10-03	799.9	n.d.	30.6	n.d.	n.d.	n.d.	n.d.	830.5
30.10-04	47.1	n.d.	n.d.	n.d.	<LOQ (5.1)	n.d.	n.d.	50.3
30.10-05	108.0	n.d.	n.d.	10.8	<LOQ (5.1)	n.d.	<LOQ (3.3)	124.2
30.10-06	371.0	4.7	24.0	n.d.	n.d.	n.d.	n.d.	399.6

*For the calculation of the total loads, ∑UV-Fs, (LOD+LOQ)/2 was taken as the concentration value for those <LOQs.

Table A8. Concentrations of UV-Fs (ng g⁻¹ lw milk) in the sample where UV-Fs were detected. <LOQ: below the limits of quantification; n.d.: not detected. In parenthesis, the value of the corresponding LOQ expressed in (ng g⁻¹ lw milk).

Sample	Concentrations ng g ⁻¹ lw							ΣUV-Fs*
	BP3	4HB	4DHB	4MBC	TBHPBT	UV320	UV329	
14.10-07	n.d.	n.d.	<LOQ	n.d.	n.d.	n.d.	n.d.	7.6
21.10-02	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	<LOQ	52.2
21.10-05	<LOQ	n.d.	n.d.	46.6	643.5	n.d.	n.d.	693.8
22.10-02	n.d.	n.d.	n.d.	n.d.	2,258.7	n.d.	n.d.	2,258.7
22.10-04	70.4	n.d.	n.d.	<LOQ	n.d.	n.d.	n.d.	96.7
22.10-06	n.d.	n.d.	n.d.	<LOQ	<LOQ	n.d.	n.d.	131.8
23.10-06	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	3.6
23.10-07	<LOQ	n.d.	n.d.	n.d.	<LOQ	n.d.	n.d.	92.1
23.10-10	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	4.5
23.10-13	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	5.1
28.10-03	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	2,050.8	2,055.0
28.10-04	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	4.6
28.10-06	n.d.	n.d.	n.d.	n.d.	<LOQ	n.d.	n.d.	102.5
28.10-08	n.d.	n.d.	n.d.	n.d.	<LOQ	n.d.	n.d.	90.7
29.10-01	n.d.	n.d.	n.d.	n.d.	<LOQ	n.d.	n.d.	57.2
29.10-06	n.d.	n.d.	n.d.	n.d.	<LOQ	n.d.	n.d.	94.3
29.10-09	975.5	n.d.	486.5	n.d.	n.d.	n.d.	n.d.	1,462.0
29.10-10	1,160.4	85.7	518.9	n.d.	n.d.	n.d.	<LOQ	1,812.9
29.10-11	1,145.3	n.d.	793.4	n.d.	n.d.	n.d.	6,661.0	5,269.2
29.10-12	6,056.2	n.d.	810.2	n.d.	n.d.	n.d.	n.d.	6,866.3
29.10-13	3,274.9	n.d.	960.1	n.d.	n.d.	7,670.7	4,812.6	21,531.0
30.10-01	1,217.6	n.d.	n.d.	n.d.	n.d.	8,376.8	2,983.2	12,577.6
30.10-02	1,348.7	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	1,348.7
30.10-03	34,183.8	n.d.	1,307.7	n.d.	n.d.	n.d.	n.d.	35,491.5
30.10-04	1,336.7	n.d.	n.d.	n.d.	<LOQ	n.d.	n.d.	1,430.4
30.10-05	2,306.6	n.d.	n.d.	230.8	<LOQ	n.d.	<LOQ	2,653.8
30.10-06	10,449.3	131.0	676.1	n.d.	n.d.	n.d.	n.d.	11,256.3

*For the estimation of the total loads, ΣUV-Fs, (LOD+LOQ)/2 was taken as the concentration value for those <LOQs. Fat values applied for each individual sample in Table A7. <LOQ not specified for each sample.

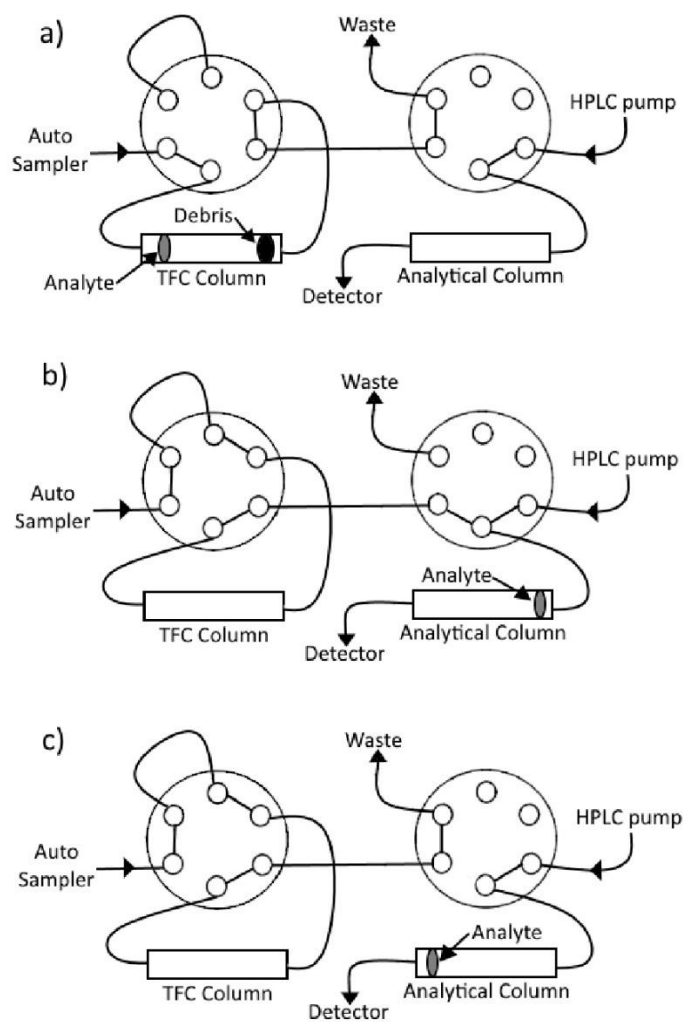


Figure A1.

1a. Focus-Mode: Loading step. The sample is injected onto the TFC column, where the analyte is retained whereas the debris from the matrix is led to waste.

1b. Focusing: Transfer step. The flow of both columns is combined and led into the analytical column.

1c. Focus-Mode: Eluting Step. The analytes eluted from the TFC column into the analytical column. The TFC column is washed and conditioned for the next injection.

